

# FOKUS methods lecture

## **CRISPR-Cas systems**

Prof. Dr. Chase Beisel

18 January, 2022

# HIRI: the first institute for RNA-based infection research



**Lars Barquist**



Integrative  
Informatics for  
Infection Biology

**Chase Beisel**



**RNA Synthetic  
Biology**

**Neva Caliskan**



Recoding  
Mechanisms  
in Infections

**Mathias Munschauer**



Viral-host  
interactions

**Emmanuel Saliba**



Single-Cell  
Analysis

**Redmond Smyth**



Genome Architecture  
and Evolution of RNA  
Viruses

**Jörg Vogel**



RNA Biology of  
Bacterial Infections

**Alexander Westermann**



Host-Pathogen-Microbiota  
Interactions

# In this lecture...

- Discovery of CRISPR
- Types and mechanisms
- Cas9 and the sgRNA

## Learning objectives

- Define CRISPR, Cas, and other basic terms
- Identify the three steps of adaptive immunity by CRISPR-Cas systems
- Explain how a CRISPR nuclease selects its target
- Explain why CRISPR-Cas systems were readily co-opted as genome-editing technologies

**What have you heard about CRISPR?**

**(Comment using chat function)**

# CRISPR for genome surgery



Home / News & Opinion

## US Companies Launch CRISPR Clinical Trial

The Germany-based study will test an ex vivo genome-editing therapy for the inherited blood disorder  $\beta$ -thalassemia.

Sep 3, 2018  
CATHERINE OFFORD

**TheScientist**  
EXPLORING LIFE, INSPIRING INNOVATION

NEWS & TECHNOLOGY 30 May 2017, updated 7 June

2017

## Boom in human gene editing as 20 CRISPR trials gear up

**NewScientist**

## Scientists Precisely Edit DNA In Human Embryos To Fix A Disease Gene

**npr**

August 2, 2017 - 1:09 PM ET  
Heard on All Things Considered

AUG 4, 2017 @ 11:37 AM 8,821

12 Stocks to Buy I

## Gene Editing Breakthrough: How Far Are We From Fixing And Designing Babies?

**Forbes**

## European court ruling raises hurdles for CRISPR crops

By Erik Stokstad | Jul. 25, 2018, 4:40 PM

**Science** MAMA

## How will we keep controversial gene drive technology in check?

By Kelly Servick | Jul. 19, 2017, 4:00 PM

**Science** MAMA

# The Nobel prize for CRISPR



2020 Nobel prize in chemistry



**Emmanuelle Charpentier**  
(Max Planck)

**Jennifer Doudna**  
(UC Berkeley)

# The mystery of CRISPR and Cas

**CRISPR** – Clustered Regularly Interspaced Short Palindromic Repeats

**Cas** – CRISPR associated



○	◆
GAGTTCCCCGCGCCAGCGGGGATAAACCG	CTTTCGCAGACGCGCGGGCGATACGCTCACGCA
GAGTTCCCCGCGCCAGCGGGGATAAACCG	CAGCCGAAGCCAAAGGTGATGCCGAACACGCT
GAGTTCCCCGCGCCAGCGGGGATAAACCG	GGCTCCCTGTTCGGTTGTAATTGATAATGTTGA
GAGTTCCCCGCGCCAGCGGGGATAAACCG	TTTGGATCGGGTCTGGAATTTCTGAGCGGTTCG
GAGTTCCCCGCGCCAGCGGGGATAAACCG	CGAATCGCGCATAACCCTGCGCGTCGCCGCCTGC
GAGTTCCCCGCGCCAGCGGGGATAAACCG	TCAGCTTTATAAAATCCGGAGATACGGAAACTA
GAGTTCCCCGCGCCAGCGGGGATAAACCG	GACTCACCCCGAAAAGAGATTGCCAGCCAGCTT
GAGTTCCCCGCGCCAGCGGGGATAAACCG	CTGCTGGAGCTGGCTGCAAGGCAAGCCGCCCA
GAGTTCCCCGCGCCAGCGGGGATAAACCG	GGGGGCGCATGACCGTAAACATTATCCCCCGG
GAGTTCCCCGCGCCAGCGGGGATAAACCG	GGAGTTCAGACATAGGTGGAATGATGGACTAC
GAGTTCCCCGCGTTAGCGGGGATAAACCG	CCCGGTAGCCAGGTTTGCAACGCCCTGAACCGA
GAGTTCCCCGCGCCAGCAGGGATAAACCG	GCAACGACGGTGAGATTTACGCCTGACGCTG
GTGTTCCCCGCATCAGCGGGGATAAACCG	GGCGCACTGGATGCGATGATGGATATCACTTG
GAGTTCCCCGCCTCTGCGG	

*E. coli* MG1655

In ~40% of bacteria, ~90% of archaea

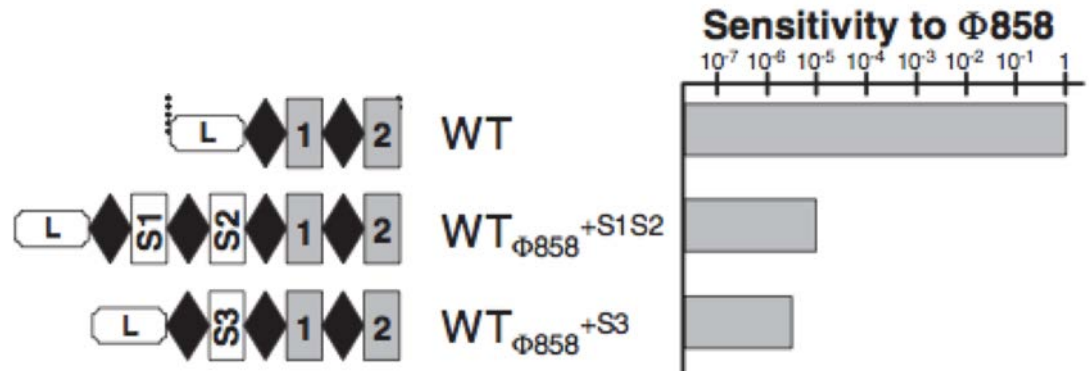
# Spacers match invader sequences

Other ID*	Species	No. of phage-matching spacers§					
		Sfi11 (AF158600)	Sfi19 (AF115102)	Sfi21 (AF115103)	DT1 (AF085222)	O1205 (U88974·1)	7201 (AF145054)
CNRZ1066	<i>S. thermophilus</i>	7	6	7	4	9	0
LMG18311	<i>S. thermophilus</i>	4	4	3	1	4	5
CNRZ302	<i>S. thermophilus</i>	2	0	0	1	2	1
CNRZ388	<i>S. thermophilus</i>	2	6	5	5	2	6
CNRZ389	<i>S. thermophilus</i>	3	2	1	2	2	5
CNRZ1100	<i>S. thermophilus</i>	2	4	2	2	2	2
CNRZ1202	<i>S. thermophilus</i>	2	5	8	4	3	2
CNRZ703	<i>S. thermophilus</i>	1	2	5	1	0	1
CNRZ1575	<i>S. thermophilus</i>	2	2	1	1	1	0
CNRZ385	<i>S. thermophilus</i>	0	3	3	2	1	3
JIM8229	<i>S. vestibularis</i>	0	0	0	0	0	0
JIM8230	<i>S. vestibularis</i>	1	1	1	0	1	0
JIM1567	<i>S. thermophilus</i>	3	4	1	1	3	2
JIM1560	<i>S. thermophilus</i>	1	1	2	0	2	0
JIM1575	<i>S. thermophilus</i>	1	1	2	0	2	0
JIM1584	<i>S. thermophilus</i>	1	1	1	1	0	0
JIM1588	<i>S. thermophilus</i>	1	1	2	0	2	0
JIM70	<i>S. thermophilus</i>	2	1	1	1	1	2
JIM71	<i>S. thermophilus</i>	1	1	1	1	0	0
JIM72	<i>S. thermophilus</i>	2	2	2	2	1	3
JIM76	<i>S. thermophilus</i>	10	8	9	6	12	0



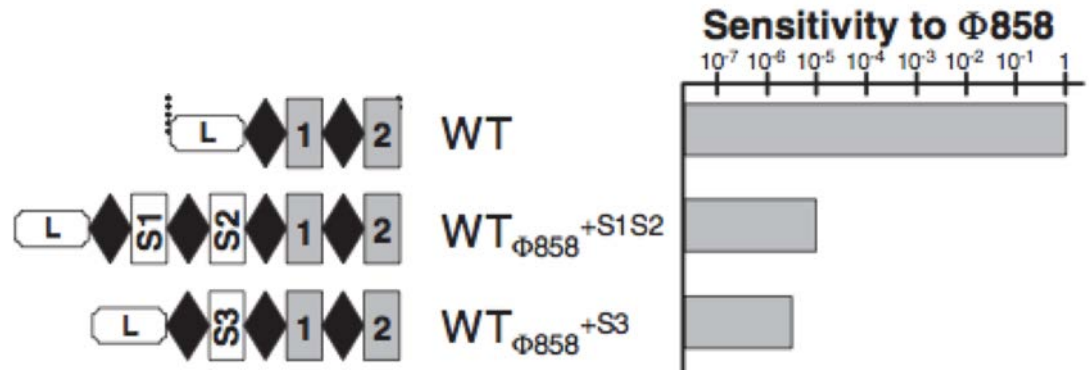
# CRISPR and Cas as adaptive defense systems

Blocks infection of  
targeted  
bacteriophages...

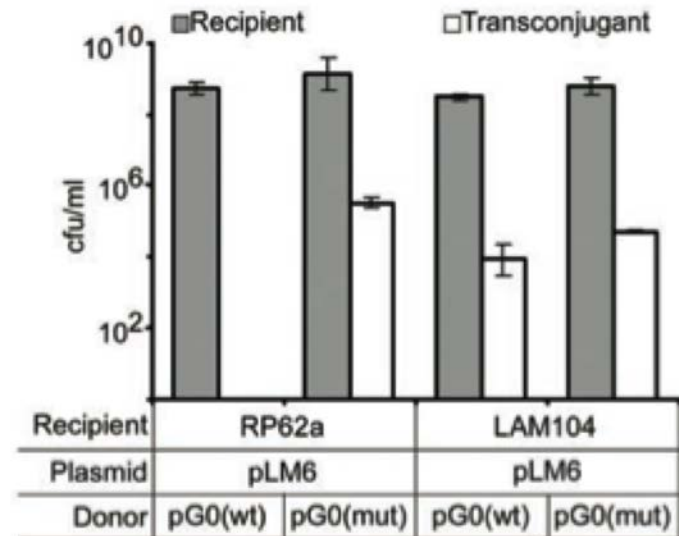


# CRISPR and Cas as adaptive defense systems

Blocks infection of targeted bacteriophages...



...and conjugated plasmids



# CRISPR and Cas as adaptive defense systems

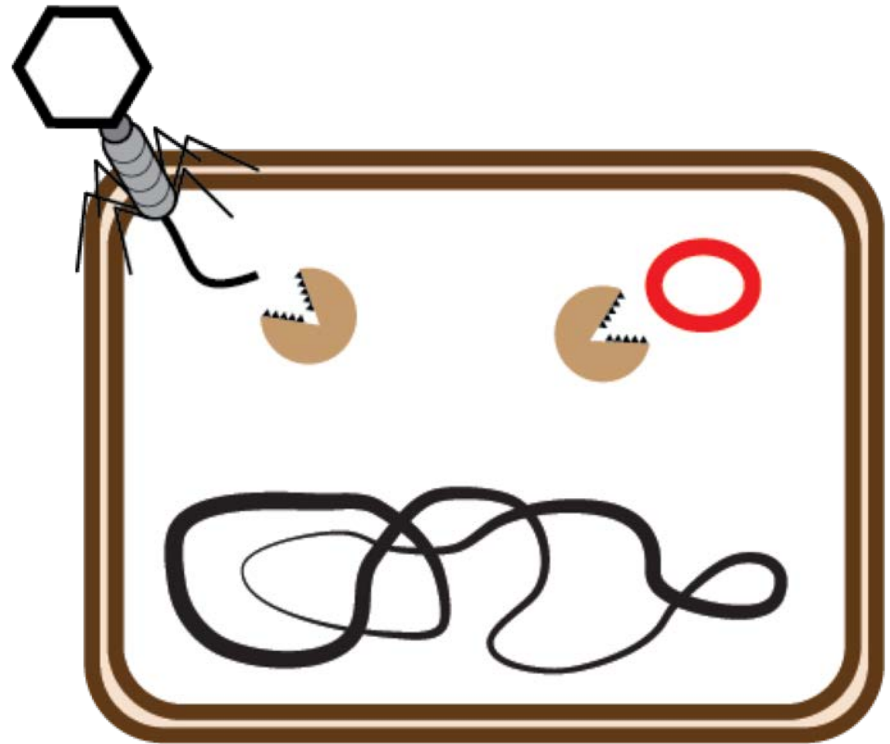
Fends off foreign invaders



Plasmids

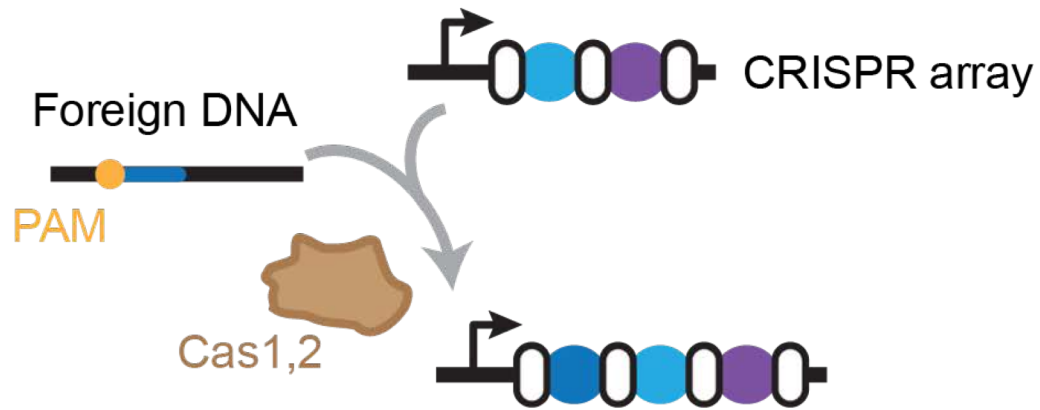


Bacteriophages



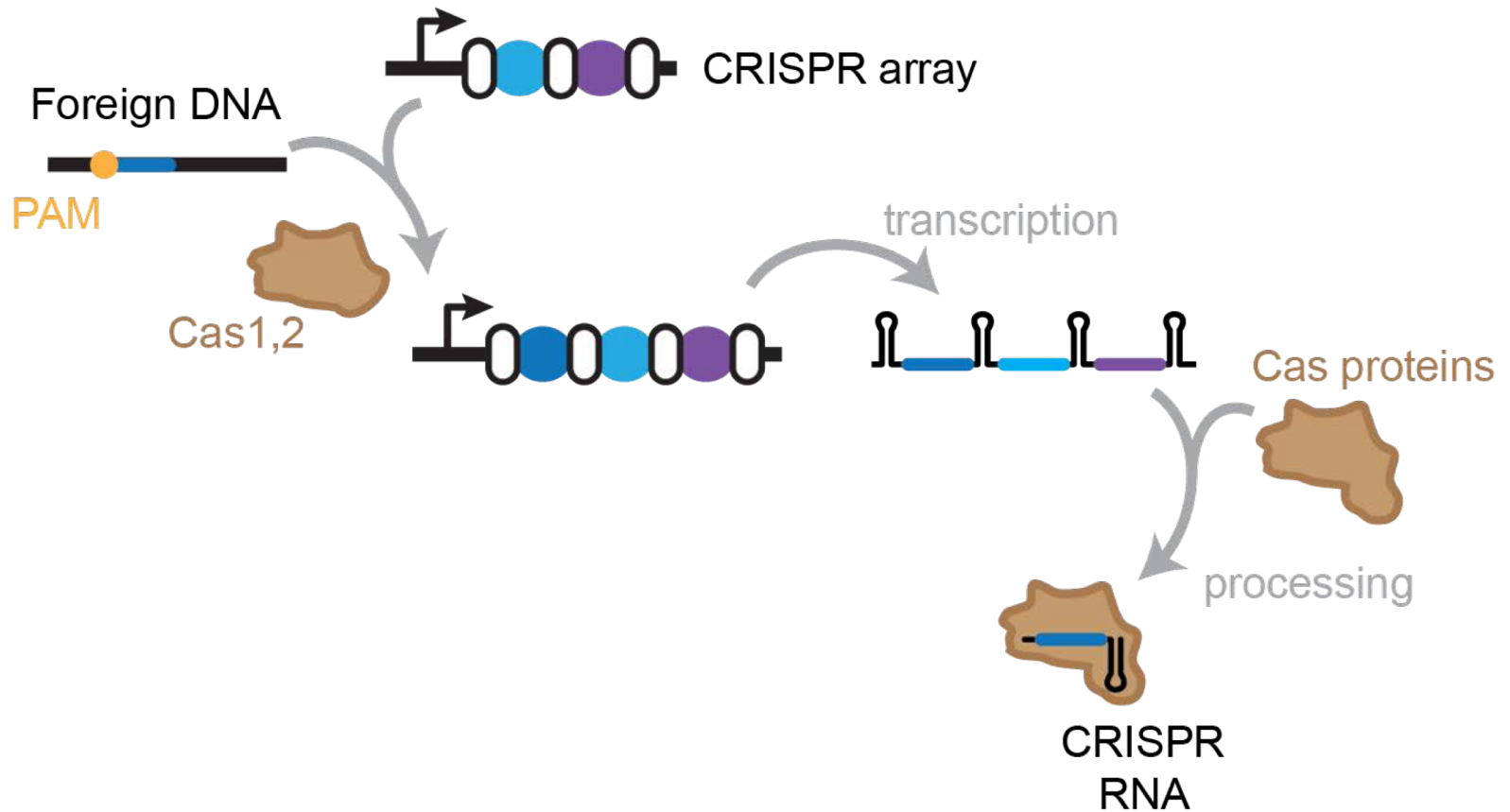
**3 steps:** Acquisition, Expression, Interference

# CRISPR and Cas as adaptive defense systems



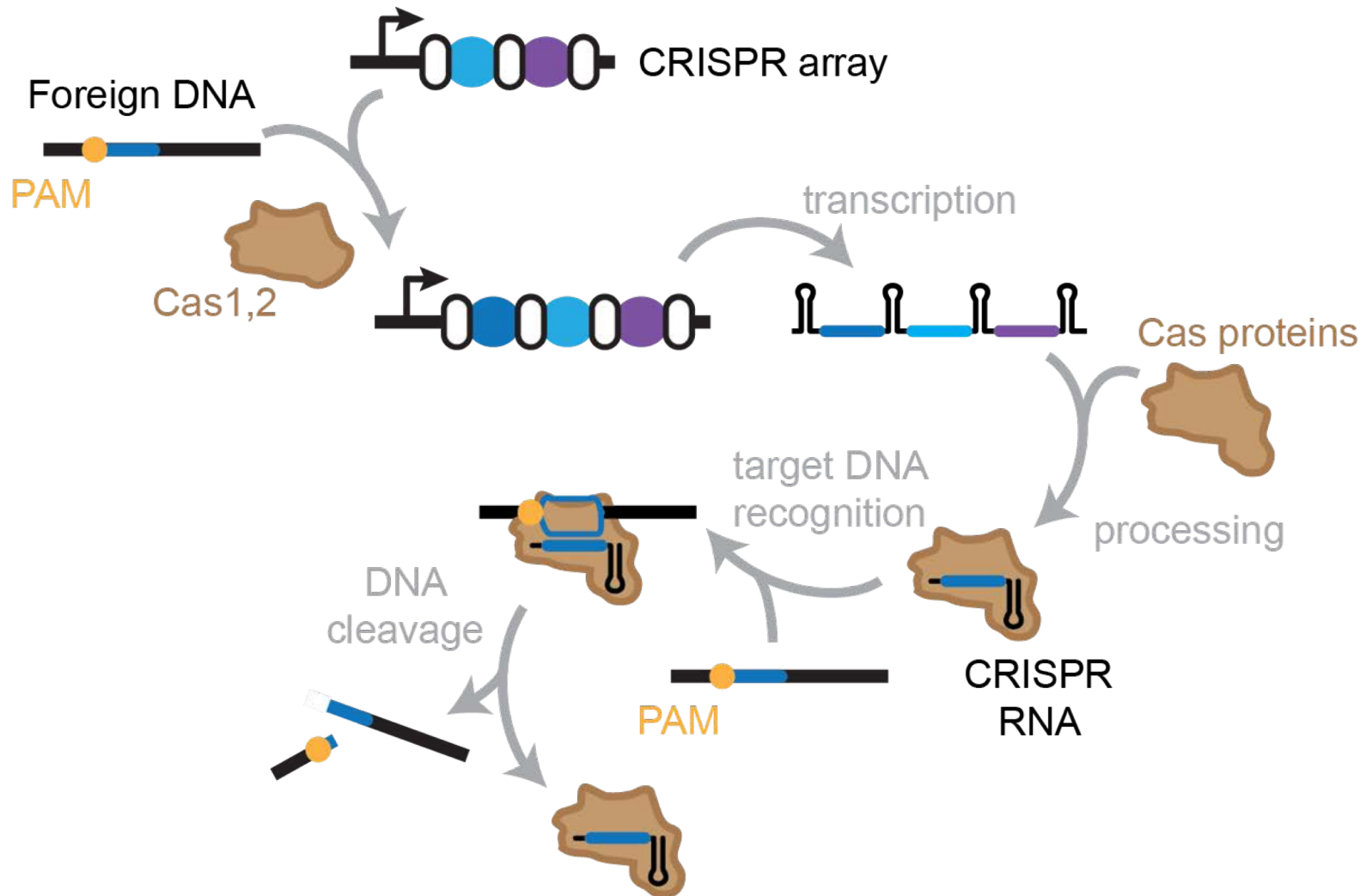
**Step 1: Acquisition**

# CRISPR and Cas as adaptive defense systems



**Step 2: Expression**

# CRISPR and Cas as adaptive defense systems



**Step 3: Interference**

# CRISPR vocabulary

**CRISPR array:** set of alternating identical repeats and distinct spacers

**CRISPR RNA (crRNA):** final, processed form of a CRISPR array

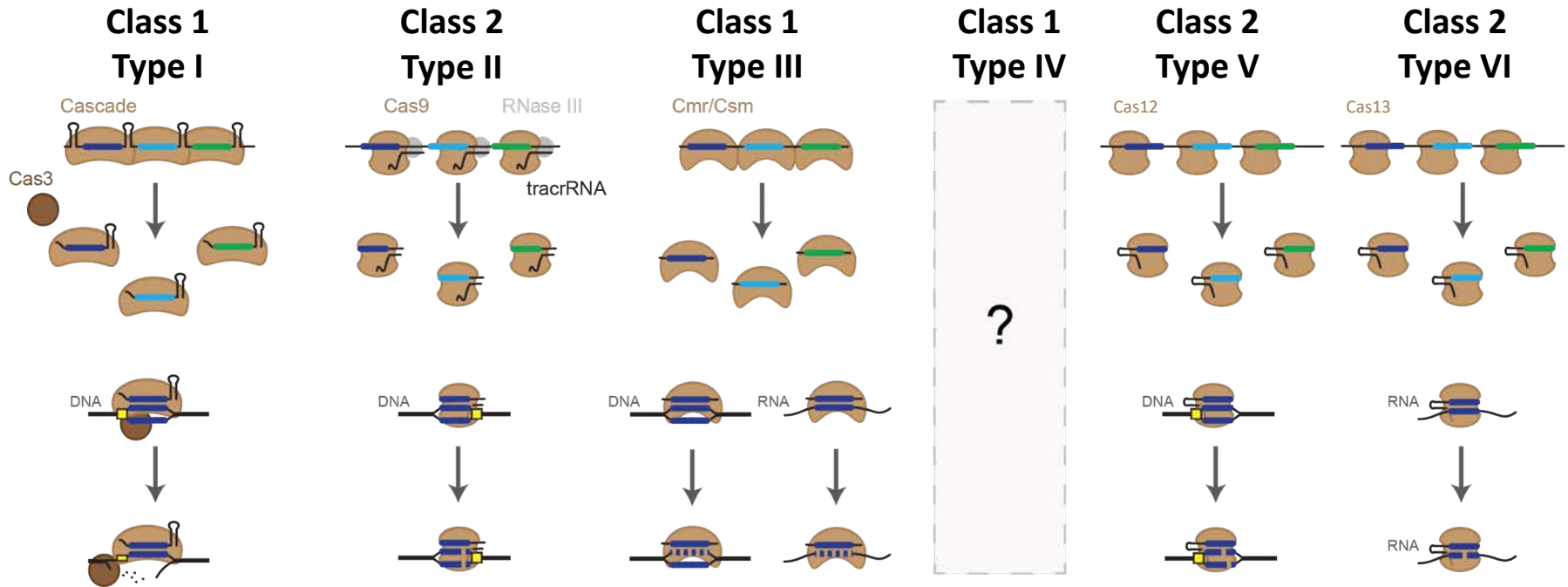
**Guide sequence:** sequence used for DNA targeting. Sometimes interchanged with spacer.

**Guide RNA:** engineered or natural CRISPR RNA

**Protospacer:** target sequence that is complementary to spacer

**PAM:** protospacer-adjacent motif, required for targeting by many CRISPR effector proteins

# Classes and types of CRISPR-Cas systems



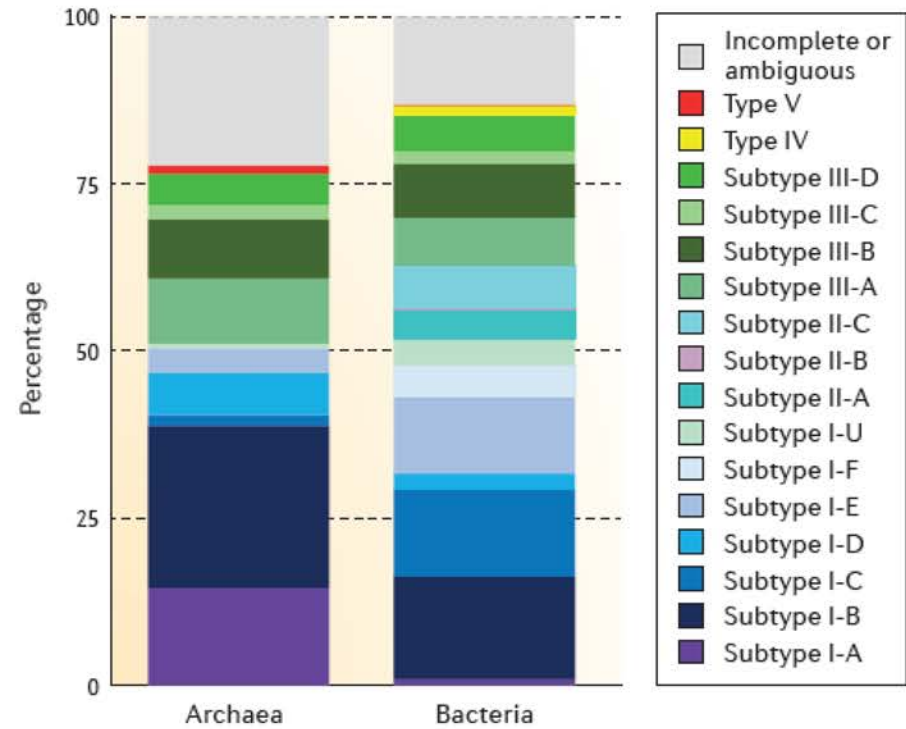
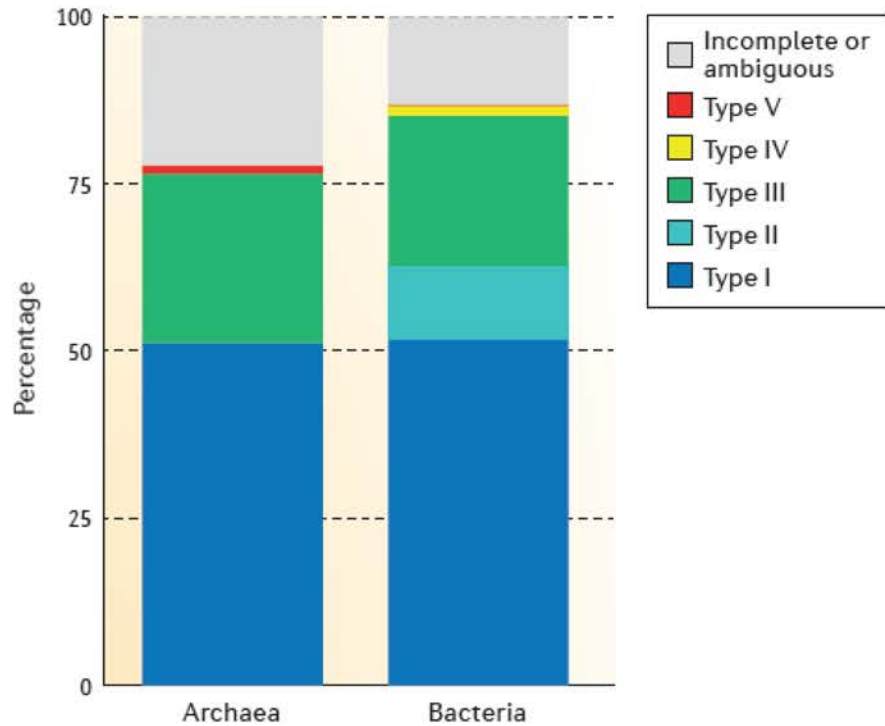
**Class (2)** – encompasses all systems with a multi-protein effector complex (**Class 1**) or single effector protein (**Class 2**)

**Type (6)** – defined by the effector complex or single-effector protein

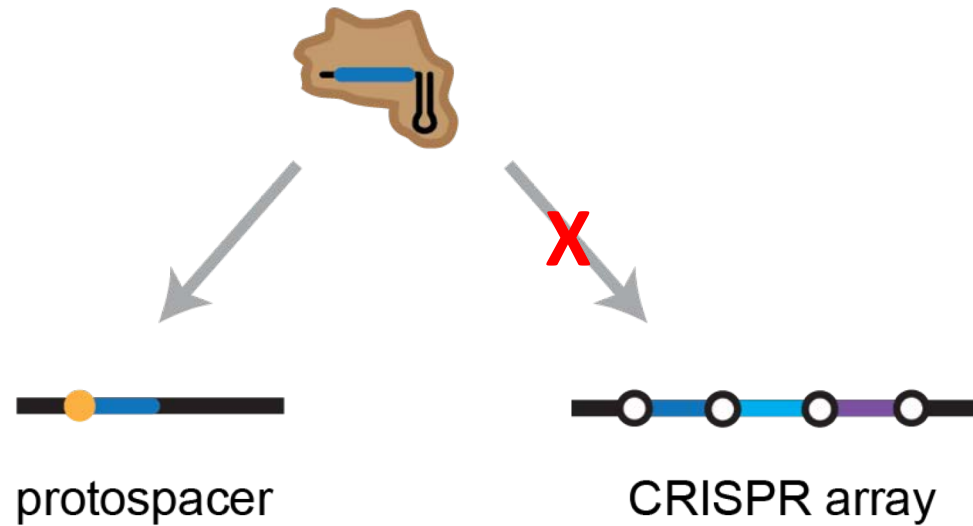
**Subtype (>30)** – defined by set and configuration of accessory proteins



# Prevalence of system types, sub-types varies widely

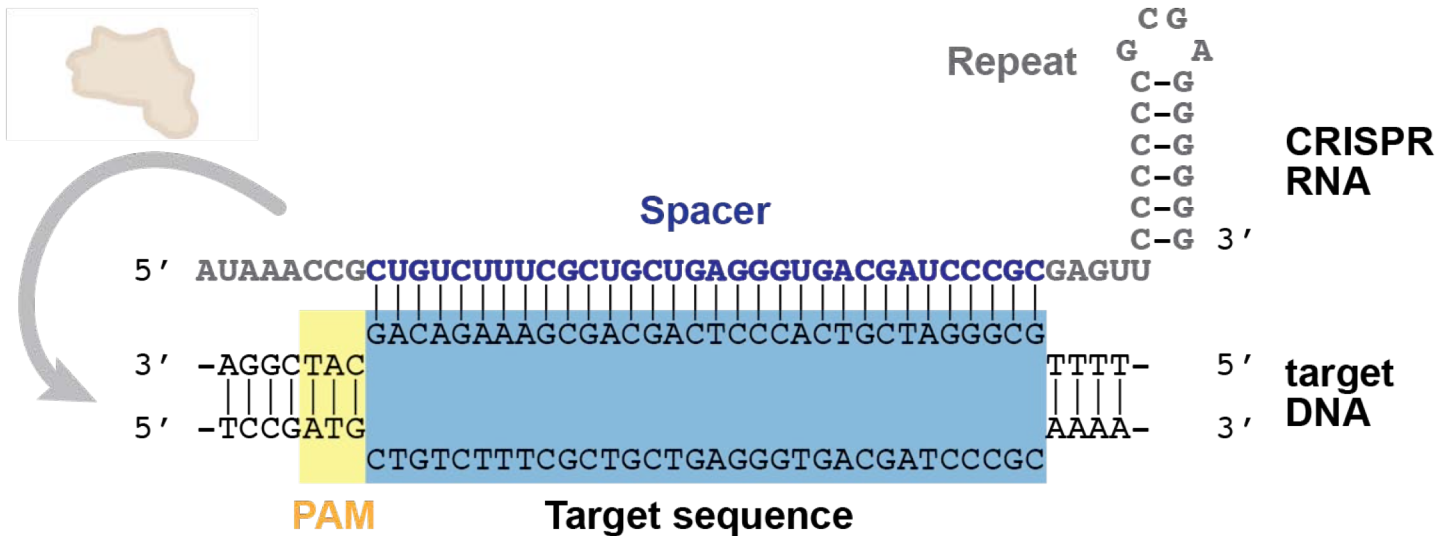


# Differentiating self from non-self



How does CRISPR recognize the target, but not its own array?

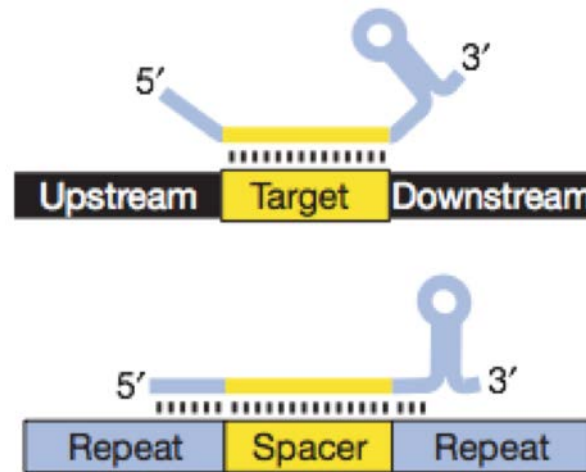
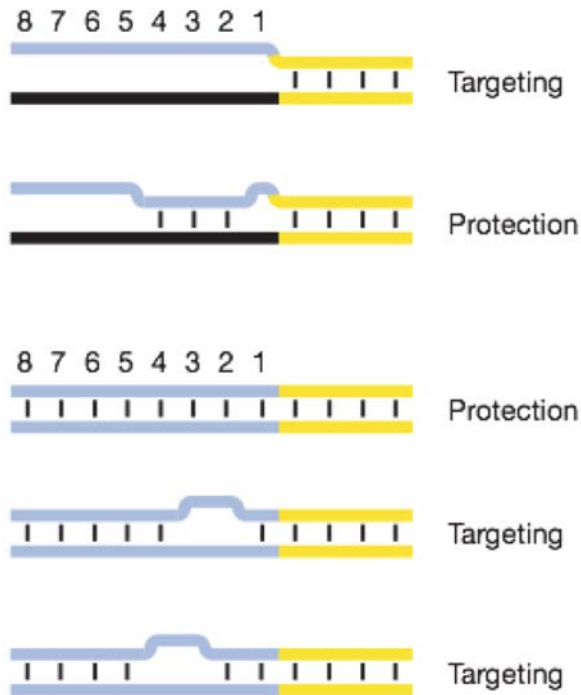
# DNA target recognition for Type I, II, IV, V systems



- PAM - protospacer-adjacent motif
- Sequence recognized by Cas protein to initiate DNA interrogation
- PAM sequence, size, and location depends on nuclease

*S. pyogenes* Cas9: **NGG PAM** on **3' end** of matching target  
*F. novicida* Cas12a: **TTTV PAM** on **5' end** of matching target

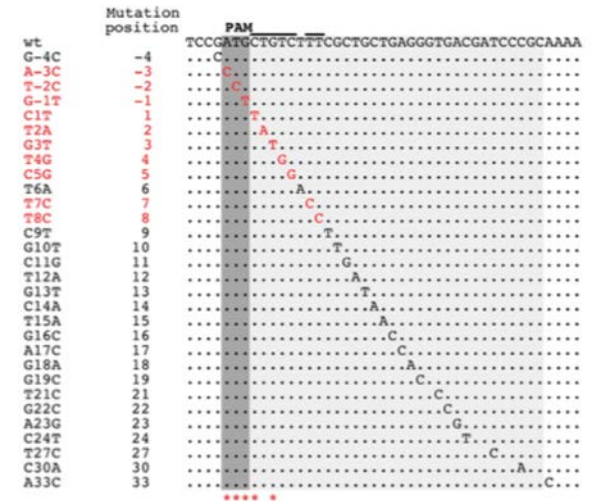
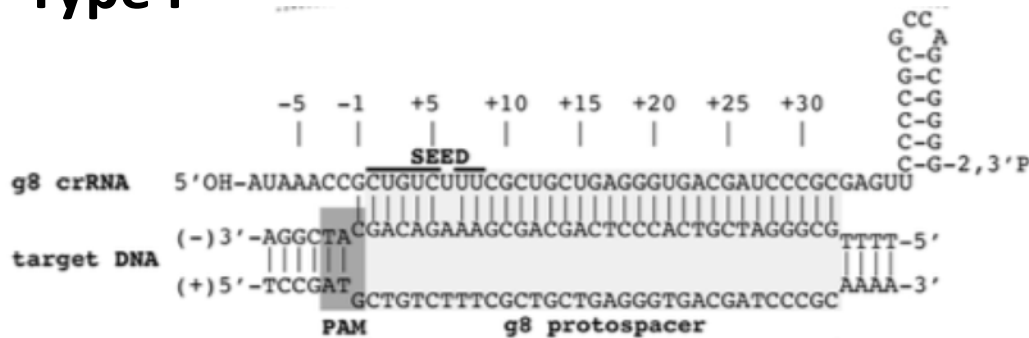
# RNA target recognition for Type III, VI systems



Marraffini & Sontheimer. *Nature* (2010)

# A spacer “seed” sequence is sensitive to mismatches

## Type I



Semenova et al. *PNAS* (2011)

- Helps ensure sequence-specificity of targeting
- Some mismatches can be accommodated, particularly outside of the seed
- Exact length, location of seed depends on the nuclease

**Why might it be advantageous for CRISPR nucleases to accept some mismatches?**

# How to identify CRISPR-Cas systems

The CRISPR database for identifying CRISPR loci and *cas* genes in sequenced prokaryotic genomes

<http://crispr.u-psud.fr>

**CRISPRs web server**

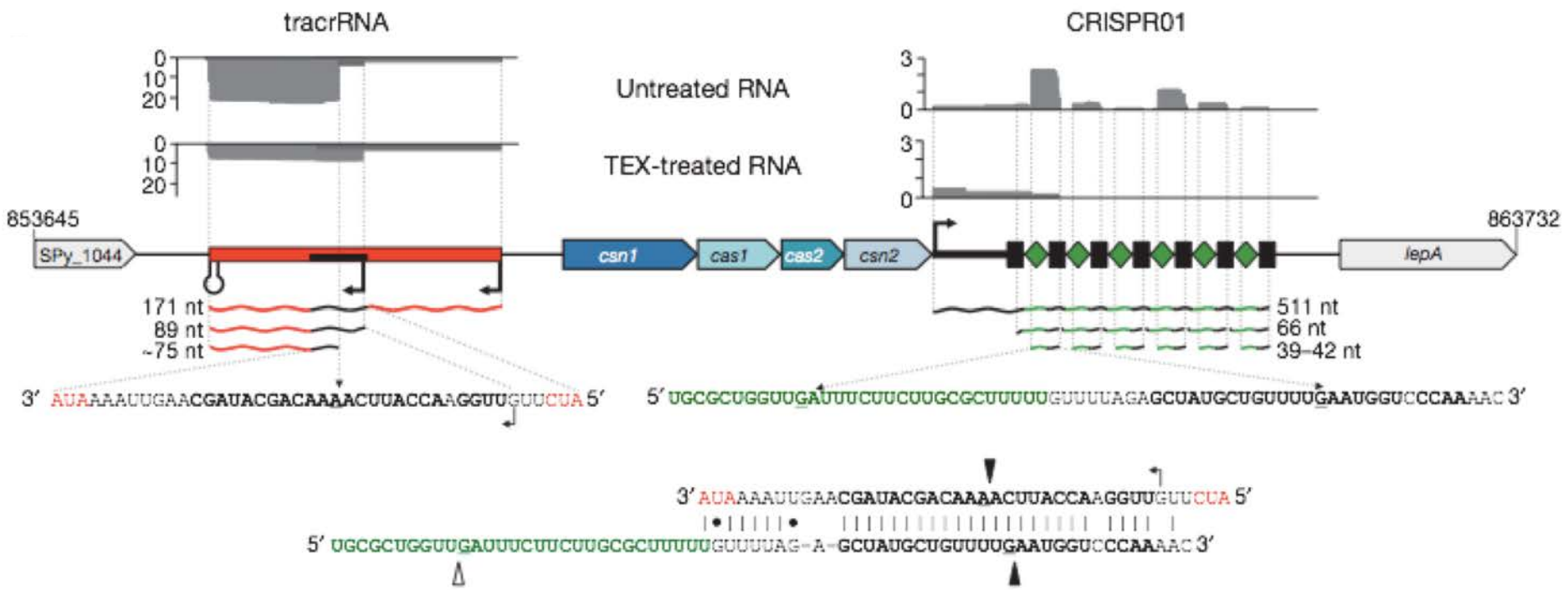
Home	About CRISPRs	News	FAQs	Help	Contact Us	Examples	IGM
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# Harnessing CRISPR

- Original studies of CRISPR focused on Type I CRISPR-Cas systems
- But Type I systems require 4 – 7 proteins in defined stoichiometries
- Type II systems require only one protein (Cas9)
- How do you generate the CRISPR RNA for Cas9?



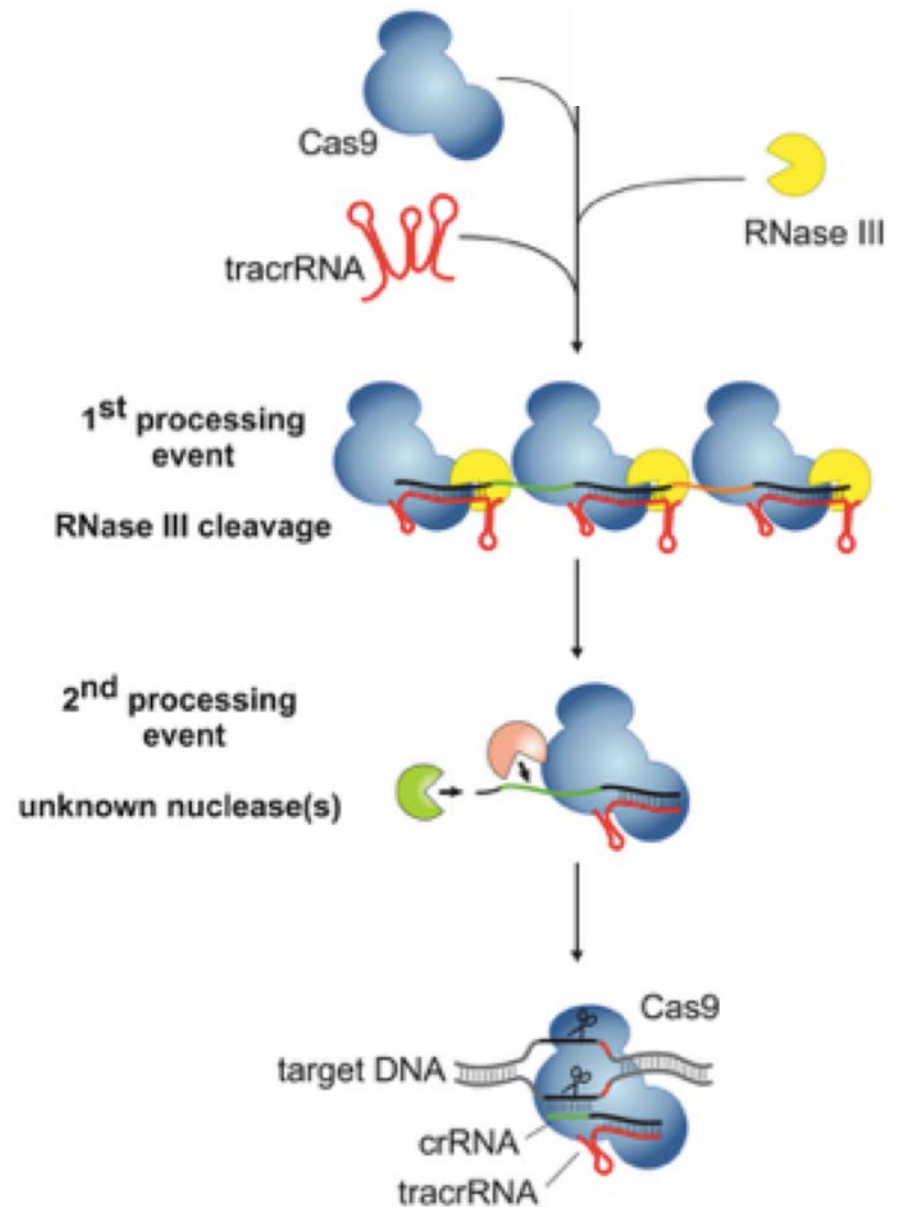
# Discovery of the tracrRNA in *S. pyogenes*



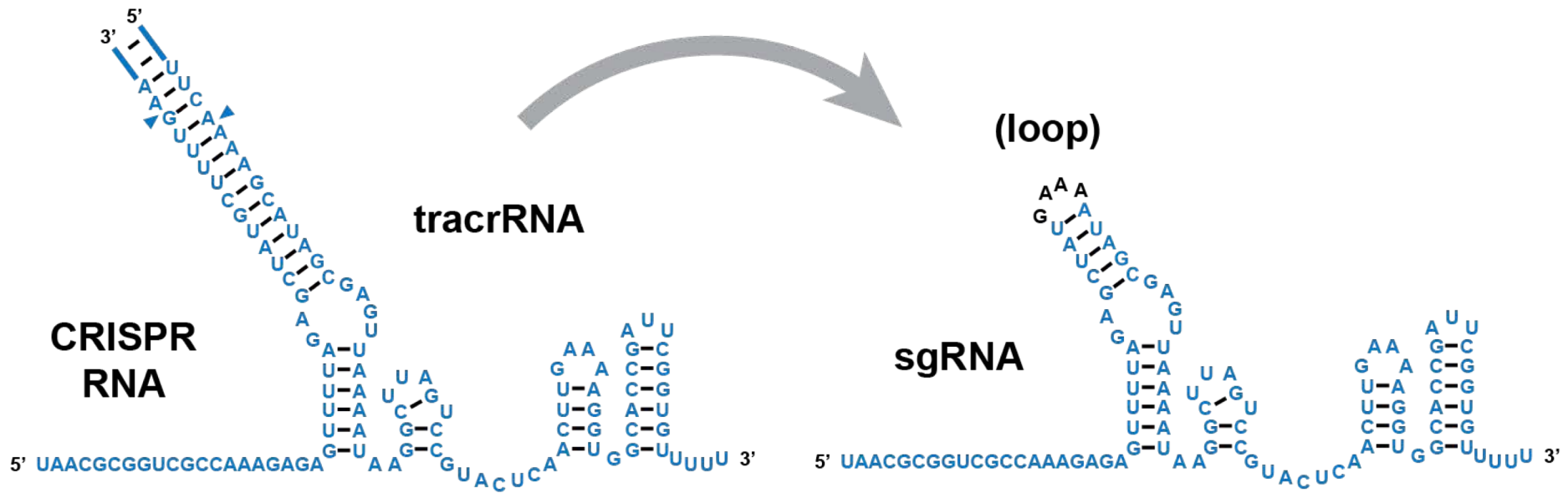
Deltcheva et al. *Nature* (2011)

# Type II Cas9 as single-effector protein with dual RNA guide

- Hybrid of crRNA and tracrRNA processed by RNase III
- 5' end of crRNA processed by unknown nucleases
- Allows for processing of array into individual crRNAs

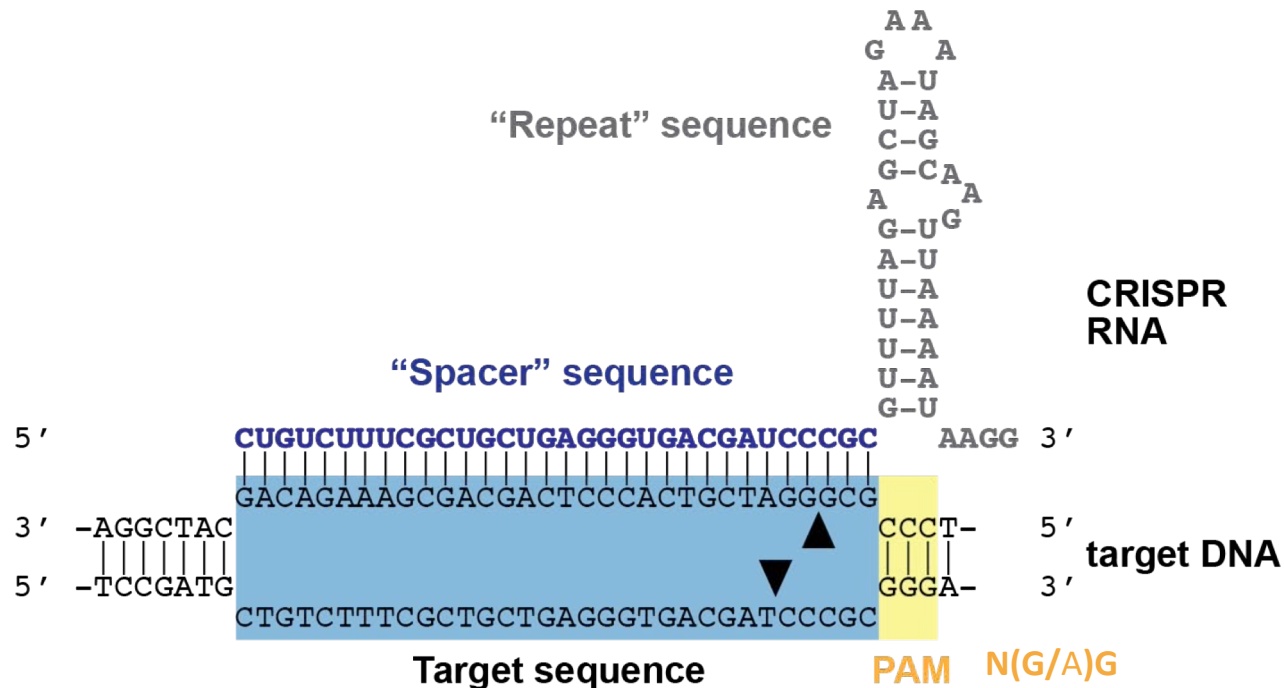


# Creating single-guide RNAs (sgRNAs)

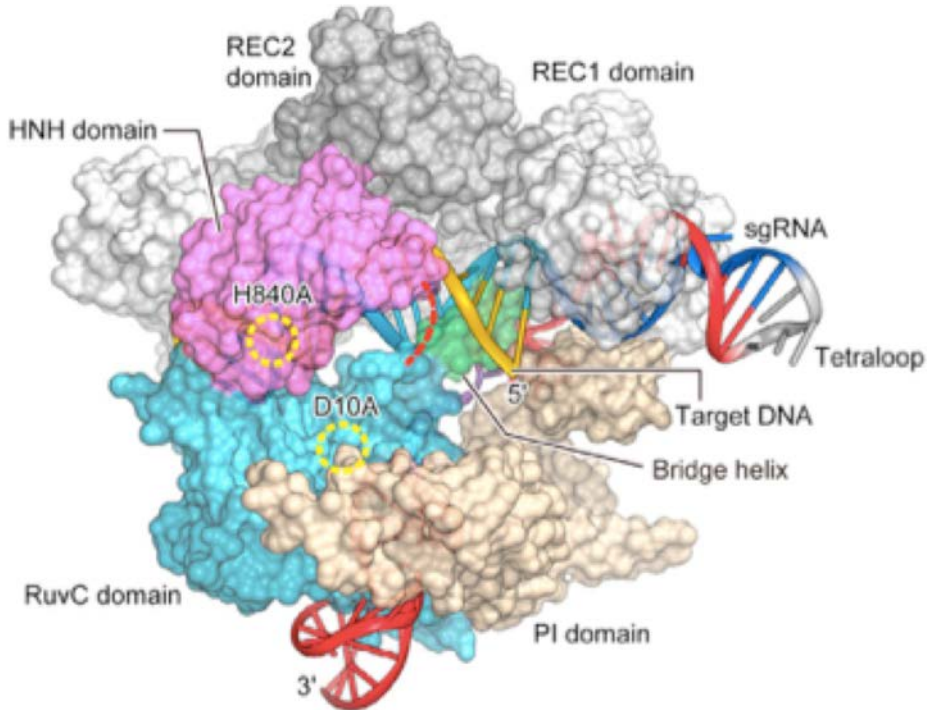


# *S. pyogenes* Cas9 as the standard

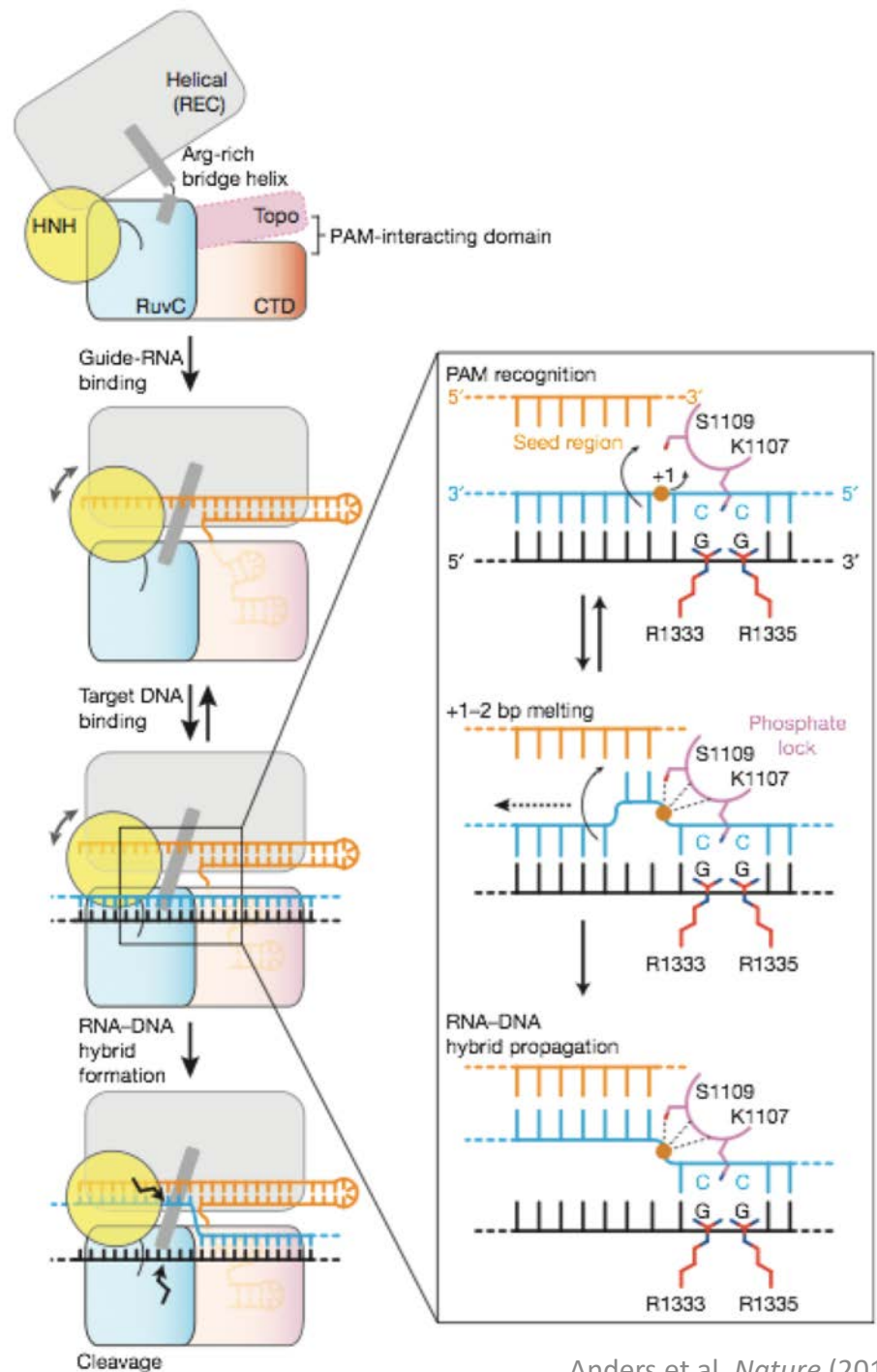
- Historical (see tracrRNA paper)
- Convenient (NGG consensus PAM)
- Numerous constructs (see Addgene)



# Cas9 as a sophisticated molecular machine

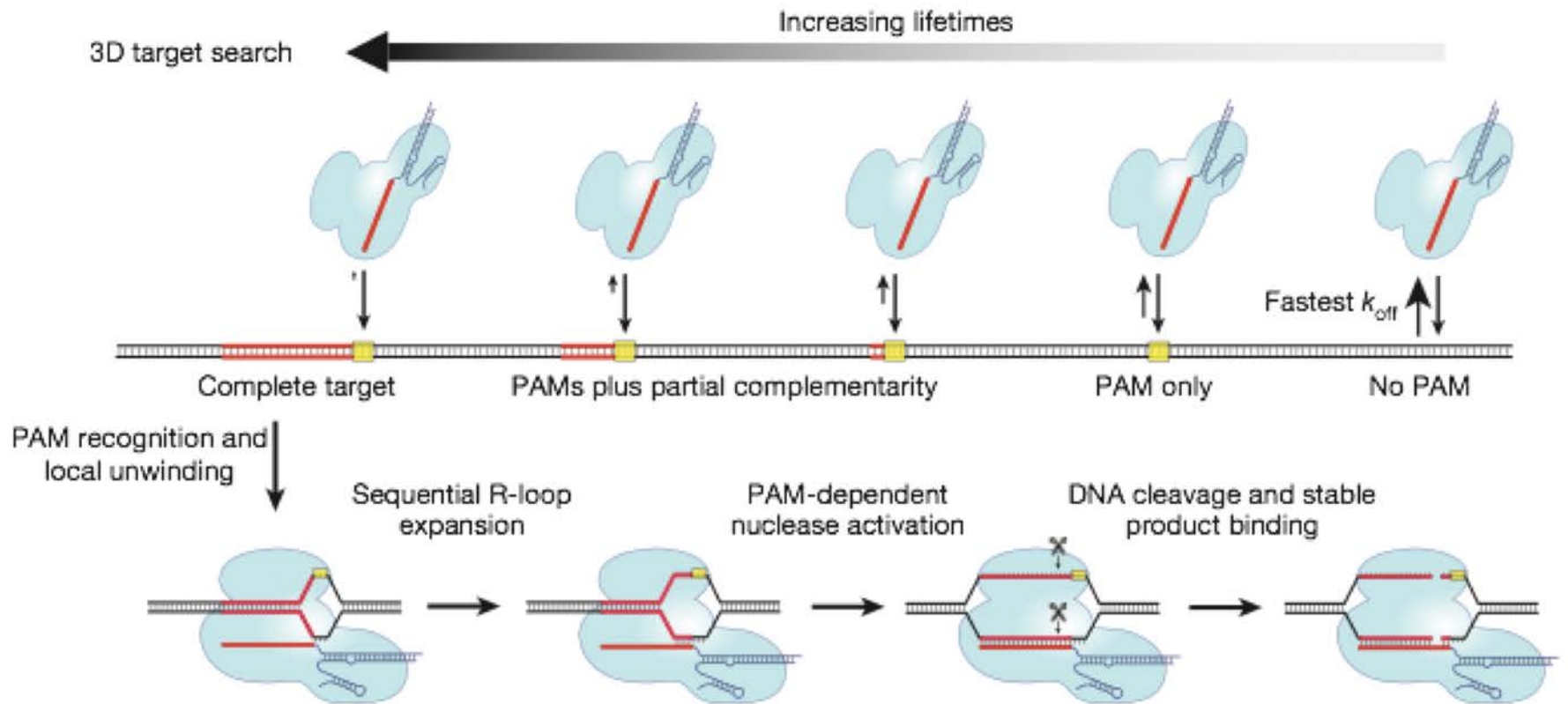


Nishimasu et al. *Cell* (2014)



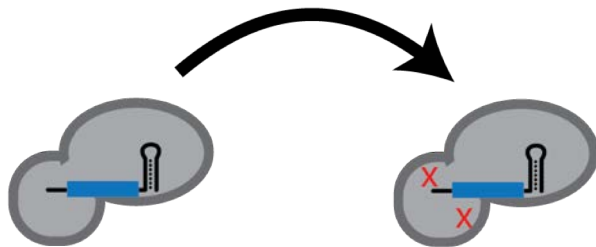
Anders et al. *Nature* (2014)

# Cas9 as a single-turnover enzyme



# Programmable binding, effector recruiting with dCas9

Introduce disruptive mutations  
in HNH, RuvC domains

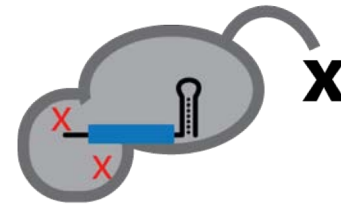


WT Cas9

dCas9

Cleaves DNA

Binds DNA



X =

- Nothing
- KRAB
- VP64
- GFP
- FokI
- APOBEC

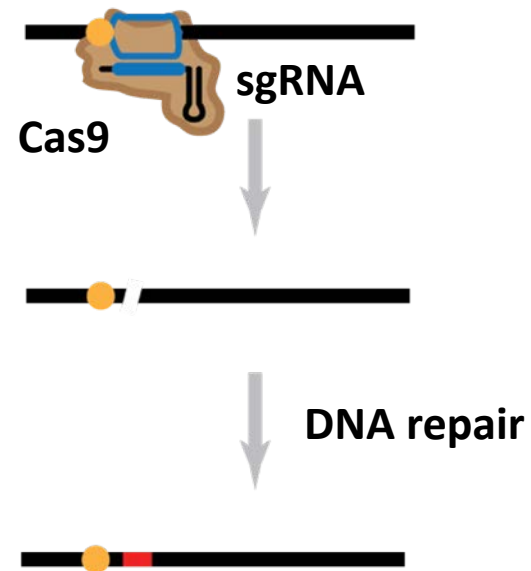
Gene regulation

Imaging

Editing

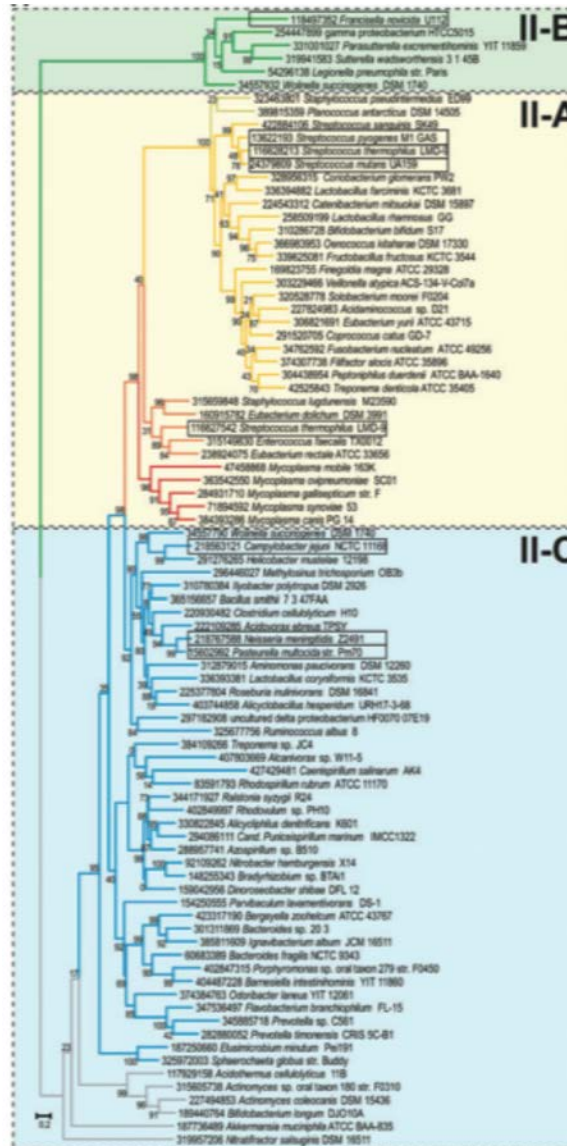
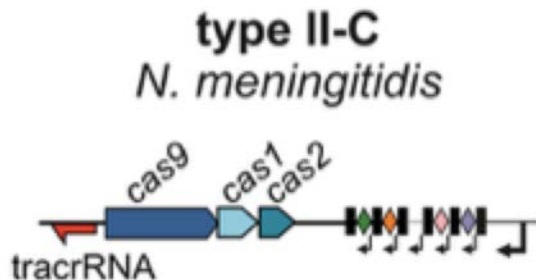
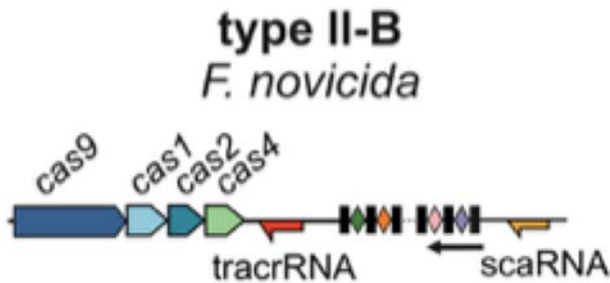
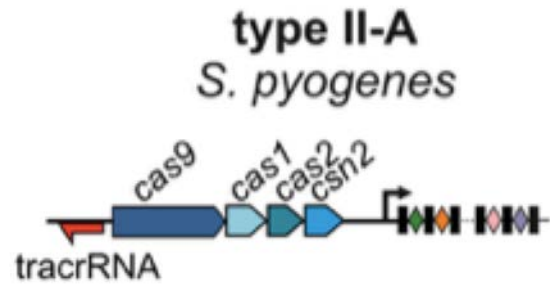
# Cas9 synonymous with CRISPR technologies

- Strain typing
- Phage resistance
- Plasmid clearance
- **Genome editing**
- Gene drives
- Gene regulation
- Antimicrobials
- Imaging
- In vitro diagnostics
- Biological recording





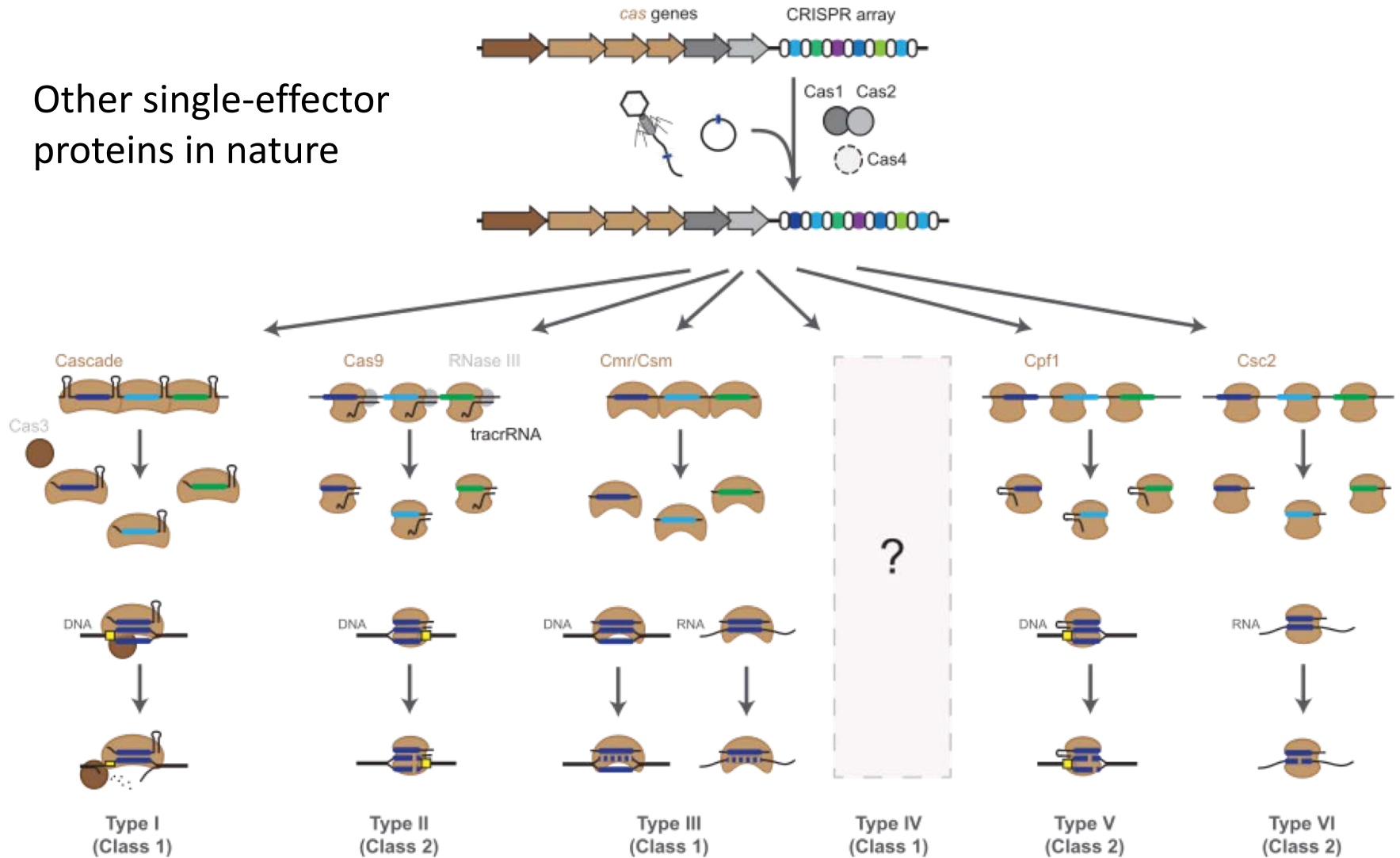
# Many other Cas9 nucleases available



- Different sizes
- Different PAMs
- Different functions
- Different immunogenicities

# Is Cas9 the best we can hope for?

Other single-effector proteins in nature



# More recently explored aspects of CRISPR

- CRISPR evolution
- Natural diversity, functions of CRISPR-Cas system
- Mechanism, application of spacer acquisition
- CRISPR transposons
- Engineering Cas proteins

# In this lecture...

- Discovery of CRISPR-Cas systems
- Types and mechanisms
- Cas9 and the sgRNA

## Learning objectives

- Define CRISPR, Cas, and other basic terms
- Identify the three steps of adaptive immunity by CRISPR-Cas systems
- Explain how a CRISPR nuclease selects its target
- Explain why CRISPR-Cas systems were readily co-opted as genome-editing technologies

**Next lecture...**

**CRISPR technologies**