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### Expanding GABA<sub>A</sub>R pharmacology via receptorassociated proteins



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#### Abstract

Drugs directly targeting  $\gamma$ -aminobutyric acid type A receptors (GABA<sub>A</sub>Rs), the major mediators of fast synaptic inhibition, contribute significantly to today's neuropharmacology. Emerging evidence establishes intracellularly GABA<sub>A</sub>R-associated proteins as the central players in determining cellular and subcellular GABAergic input sites, thereby providing pharmacological opportunities to affect distinct receptor populations and address discrete neuronal dysfunctions. Here, we report on recently studied GABA<sub>A</sub>R-associated proteins and highlight challenges and newly available methods for their functional and physical mapping. We anticipate these efforts to contribute to decipher the complexity of GABAergic signalling in the brain and eventually enable therapeutic avenues for, so far, untreatable neuronal disorders and diseases.

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### Introduction

 $\gamma$ -Aminobutyric acid type A receptors (GABA<sub>A</sub>Rs) are ion channels, gated by GABA, the major inhibitory neurotransmitter in the human brain. GABA<sub>A</sub>R subtypes are heteropentamers assembled from 20 subunits. GABA<sub>A</sub>R subtype populations exhibit specific cellular and subcellular distributions, are activity-dependently regulated during development [1] and altered in response to substances [2] and pathological conditions

such as neurodegeneration [3]. The impressive functional heterogeneity of GABAergic synapses is primarily determined by presynaptic input, local translation [4], alternate splicing of transcripts, and intracellular receptor interactions [5]. Receptor surface numbers, subcellular distribution, nanoscale arrangement [6-10], and turnover are determined by distinct and subunitspecific intracellular interactions with associated proteins, many of which are tightly regulated by posttranslational modifications [11,12]. Defects in these intracellular GABAAR signalling pathways result in GABAergic synapse dysfunction and are at the core of a number of neuropathological conditions including acute brain disorders, neuropsychiatric and neurodegenerative diseases [13] as well as neurodevelopmental disorders [14] and epileptogenesis [15]. Resolving the underlying, largely unexplored, interfaces between GABAARs and their associated proteins addresses a key challenge in neuroscience, namely linking the activity of GABAAR populations at specific input sites and within defined circuits to behavior and has great potential to expand GABA<sub>A</sub>R pharmacology [12] (Figure 1).

Here, we report on new insights into GABA<sub>A</sub>R accessory proteins and their implications for GABA<sub>A</sub>R pharmacology. Furthermore, we highlight newly available methodological approaches to identify and study the relevant interaction interfaces of the rapidly increasing number of putative GABA<sub>A</sub>R-associated proteins, —thereby provide a toolbox to unleash the full neuropharmacological potential of GABA<sub>A</sub>R-associated proteins and decipher the function of discrete GABA<sub>A</sub>R populations and specific GABAergic input sites.

# Expanding GABA<sub>A</sub>R pharmacology via the associated proteins

### Intracellularly associated proteins provide bidirectional control over GABAergic activity

Our insight into the molecular mechanisms that determine the diversity of GABA<sub>A</sub>Rs at inhibitory postsynaptic densities remains largely incomplete, even more so, their contribution to mechanisms of postsynaptic plasticity [16]. GABA<sub>A</sub>R-associated proteins that act as scaffolds or clustering factors are often membrane-associated via inositol-phosphate binding or palmitoylation [17,18]. By providing receptor-binding sites with varying affinity, density, and subsynaptic





**Targeting specific GABAergic input regions via GABA<sub>A</sub>R-associated proteins**. Schematic representation of a pentameric GABA<sub>A</sub>R bound to associated proteins that determine the clustering, half-life, and nanoscale arrangement of receptor populations. Orthosteric and allosteric effectors adjust function of GABA<sub>A</sub>R subtypes distributed across brain regions and neuronal substructures (white circles). Conceptually, pharmaceutical interference with intracellular GABA<sub>A</sub>R interactors could provide the means to modulate GABAergic signalling in specific brain regions, circuits, and even GABAergic inputs (colored circles).

arrangement [6–10], they dynamically recruit GABA<sub>A</sub>R subtypes to distinct, post-, extra-, or subsynaptic domains or regions (Figure 2) [19,20]. Compounds targeting these binding sites diminish the synaptic clustering of GABA<sub>A</sub>Rs, resulting in reduced amplitude and frequency of miniature inhibitory postsynaptic currents (mIPSCs) [21,22].

GABA<sub>A</sub>R-associated proteins mediating receptor trafficking typically control receptor numbers and half-life at the neuronal surface membrane. Direct competition with binding of the clathrin adaptor protein 2 complex (AP2) with GABA<sub>A</sub>Rs rapidly affects mouse behavior by increasing synaptic GABAAR numbers, inhibitory synaptic strength and neuronal excitability [23] (Figure 2). Recent transgenic proteomic analysis identified Cleft lip and palate transmembrane protein 1 (Clptm1) as a novel GABA<sub>A</sub>R-associated protein that limits the forward trafficking of a number of GABAAR subtypes via direct interactions [24]. The sorting nexin 27 protein (SNX27) enhances the membrane retrieval of the inhibitory synapse-specific adhesion molecule neuroligin 2 (NL2) by direct binding via its postsynaptic density-95, discslarge, zona occludens 1 (PDZ) domain [25]. The observed enhance in postsynaptic gephyrin and GABAAR clustering and consequently increased inhibitory currents upon knock-down were proposed to result from the stabilization of extrasynaptic GABA<sub>A</sub>Rs [25]. An additional, recent example is the cytoplasmic FMR1 interacting protein 1 (CYFIP1), its conditional loss alters neuroligin 3 (NL3) and GABA<sub>A</sub>R  $\beta$ -subunit expression and thereby also increases inhibitory synaptic clusters and hence amplitude of mIPSCs [26].

Their pharmacological targeting can be expected to provide antagonising and bidirectional control over GABAergic signalling, potentially enabling precise enhancement or reduction of GABAergic inhibition to address individual synaptic dysfunctions (Figures 1 and 2).

### The full potential of GABA<sub>A</sub>R subtype-specific modulation remains largely untapped

Benzodiazepines [27–29] and increasingly neurosteroids [30] contribute significantly to today's neuropharmacology. Side-effects and recognition of functional GABA<sub>A</sub>R subtype heterogeneity encouraged the development of modulators with improved subtype selectivity. Robust preclinical data recently validated the long-standing  $\alpha 2/\alpha 3$ -GABA<sub>A</sub>R positive allosteric modulator nonsedating anxiolytic and the  $\alpha 5$ -GABA<sub>A</sub>R negative allosteric modulator cognition enhancer hypotheses





**GABA<sub>A</sub>R populations are in dynamic equilibrium** [6]. GABA<sub>A</sub>R-associated proteins enrich GABA<sub>A</sub>Rs at extra- and postsynaptic sites and mediate endocytosis by interaction with specific subunits. Surface representation of GABA<sub>A</sub>R-associated proteins in blue together with known or putative binding motifs shown as red stick models. Gephyrin mediates synaptic clustering of  $\alpha$ 3-GABA<sub>A</sub>Rs via direct interaction and inhibition affects GABA<sub>A</sub>R clustering and fast synaptic inhibition [21,22]. Conversely, radixin clusters extrasynaptic pools of  $\alpha$ 5-GABA<sub>A</sub>Rs [53]. AP2 controls GABA<sub>A</sub>R surface numbers of  $\gamma$ 2-GABA<sub>A</sub>Rs [23] and  $\beta$ 3-GABA<sub>A</sub>Rs [72]. Collybistin maintains  $\alpha$ 2-GABA<sub>A</sub>Rs at the AIS of cortical pyramidal neurons [45]. Enhanced targeting  $\alpha$ 1-GABA<sub>A</sub>Rs provide seizure resilience [46]. Ankyrin/spectrin complexes are a major component of the AIS scaffold and contribute to GABA<sub>A</sub>R maintenance.

[27], thus providing the long anticipated starting points for the development of innovative treatments of cognitive disorders and anxiety and possibly even stroke, schizophrenia, and depression [27–29]. So far, the development of disease-specific allosteric GABA<sub>A</sub>Rdirected drugs has suffered from innate structural limitations of the druggable orthosteric and allosteric GABA<sub>A</sub>R sites, especially in the case of compounds designed to differentiate between  $\alpha 1$ -,  $\alpha 2$ -, and  $\alpha 3$ -GABA<sub>A</sub>Rs [27,28]. All extracellularly binding compounds, including benzodiazepines [27–29], z-drugs [31], and dihydromyricetin [32], some of them claimed to be selective for a specific receptor subtype, exhibit only limited selectivity, which is even further aggravated by their high potency [29].

GABA<sub>A</sub>R subunit-specific intracellular phosphorylation was recently associated with the impressive long-term therapeutic action of the transmembrane-binding neurosteroid Zulresso® [33]. This compound, just like the newly developed, orally bioavailable Zuranolone [34], however, acts across receptor subtypes, thus motivating alternative approaches to achieve selective GABA<sub>A</sub>R modulation.

### Targeting specific neuronal types and circuits via $GABA_AR$ -associated proteins

Intracellular interactions of GABA<sub>A</sub>Rs are highly subtype selective [12] and provide a steadily increasing (Table 1) number of binding pockets for affecting distinct receptor subtype populations. Targeting GABAAR protein complexes, instead of directly adjusting gating or conductance of a single receptor subtype across all brain regions and substructures, may provide the means to affect disease relevant protein complexes in distinct brain regions and circuits and possibly even specific neuronal types and substructures (Figure 1). This invaluable specificity advantage enables the development of sophisticated tools to explore the underlying mechanisms of GABAAR subtype distribution and maintenance. In this framework, specific micro- and macrocircuits and their corresponding behavioral outputs may be selectively modulated. This pharmacological approach can be alternative or additive to future genetic interventions for the cure of currently untreatable neurological diseases and disorders. Acute modulators of intracellular GABAAR interactions are an attractive alternative to genetic approaches, that suffer from poor correlation of functional effects to individual receptor subtype populations, as exemplified by the complex alterations in the GABA<sub>A</sub>R  $\alpha$ 1 knockout (KO) mouse brains [35]. Besides overcoming the adaptive responses in GABAergic signalling in vivo, acute modulators offer a largely improved temporal resolution and the possibility to tune effects by adjusting the dose. Recently developed chemicogenetic approaches combine acute pharmacology with genetics to achieve

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Associated protein	Mode of action	Experimental model	Binding motif
SNX27	Endocytotic recycling of NL2 at GABAergic synapses	Hippocampal neurons [25]	PDZ-mediated binding
CYFIP1	Bidirectional control over NL3 and β2/3-GABA <sub>A</sub> R subunit expression	Hippocampal neurons [26]	not defined
Radixin	Clustering of extrasynaptic $\alpha$ 5-GABA <sub>A</sub> Rs	Hippocampal neurons and KO mouse model [53], phage display and computational modeling [68]	<sup>342</sup> NYFTKRGWAWDGKKAL <sup>357</sup>
Collybistin	Clustering of postsynaptic $\alpha$ 2-GABA <sub>A</sub> Rs	GABA <sub>A</sub> R α1/2 chimera mouse model [45,46]	<sup>360</sup> VMIQNNAYAVAVANYA <sup>375</sup>
Gephyrin	Clustering of postsynaptic $\alpha$ 1/3-GABA <sub>A</sub> Rs	GABA <sub>A</sub> R α1/2 chimera mouse model [45,46], anxiety mouse model [32]	<sup>360</sup> LIKKNNTYAPTATSYT <sup>375</sup> <sup>395</sup> FNIVGTTYPIN <sup>405</sup>
NrCAM	Complex formation with synaptic scaffolding protein gephyrin via astrocytic input	In vivo Split-TurboID [48]	Gephyrin-mediated binding
Clptm1	Limiting the forward trafficking of GABA <sub>A</sub> Rs with differing subunit composition	Transgenic proteomic analysis [24]	not defined

cell-specific control over neuronal signalling proteins, but their complexity hampers broader application [36].

Targeting the associated proteins also provides a new perspective to counter the loss of GABA<sub>A</sub>Rs and their postsynaptic scaffold in benzodiazepine pharmacoresistance [37]. Disrupting or stabilizing scaffolding/trafficking interactions could help to overcome tolerance and reduce dose escalation, which is urgently required for benzodiazepines that are broadly prescribed for the treatment of anxiety and insomnia [38].

### Reconstitution of neuronal dysfunction via GABA<sub>A</sub>Rassociated gephyrin and collybistin

Gephyrin and collybistin are critical for the specification and maintenance of soma and axon targeting GABAergic contacts in the cortex and hippocampus that are implicated in epileptic disorders [39,40]. Gephyrin mediates the alternative recruitment of distinct GABAAR and glycine receptor subunits via a universal receptor binding pocket [41]. Upon receptor binding and regulated by post-translational modification, gephyrin forms a highly condensed sheet-like assembly via phase separation [42]. Competition with receptor binding modulates fast synaptic inhibition by reducing clustering and thus conductivity of a distinct subset of GABAAR subtypes. This was initially achieved using native fragments of GABA<sub>A</sub>R and glycine receptors [41], later with affinity enhanced variants [21,43] and recently even small molecules [22].

Collybistin plays a significant role in the regulation [44] of the GABAergic postsynapse and enriches  $\alpha 2$ -

GABAARs at the axon-initial-segment (AIS) of cortical pyramidal cells [45]. Its dysfunction has been linked to hyperekplexia, epilepsy, intellectual disability, and developmental disorders. Mutation of the a2-GABAAR binding region of collybistin to the  $\alpha$ 1-GABA<sub>A</sub>R sequence results in loss of a distinct subset of inhibitory synapses and increased susceptibility to seizures, thus reproducing the effects of human epileptogenic collybistin mutations [45]. The observation that the seizure susceptibility could be rescued by treatment with AZD7325, an a2/3-selective positive allosteric GABAAR modulator, suggests a value of this compound for treating a subgroup of epilepsy patients. In a follow-up study, where  $\alpha$ 1-GABA<sub>A</sub>R was mutated to contain the  $\alpha$ 2 subunit binding region of collybistin, Nathanson et al. generated mice that showed a robust resilience to kainate induced seizures. Notably, these mutations were sufficient to fully rescue the lethal seizure phenotype of the existing  $\alpha 2/\alpha 1$  chimera mice [46]. Thus, substantiating that specific dysfunctional neuronal pathways can indeed be reconstituted via the intracellularly associated proteins.

### Targeting specific GABAergic input regions via $GABA_AR$ -associated proteins

Most GABAergic contacts are formed onto postsynaptic dendrites, including spines. Compared with soma and AIS, dendritic inhibition is overwhelmingly represented and targeting this key element of GABAergic inhibition blocks important forms of synaptic plasticity [4,16]. The complexity of GABAergic signalling is further increased by the recent observation that dominant GlyR  $\alpha$ 1 mutants assemble into pre- and extrasynaptic GABA<sub>A</sub>Rs [47] and that GABAergic inputs also encompass astrocyte-neuron junctions, stabilized via transcellular coupling of neuronal cell adhesion molecules to gephyrin [48].

The AIS is a crucial subcellular domain in neurons where action potentials are initiated and plays a pivotal role in neuronal physiology. The enrichment of distinct GABAAR subtypes to the AIS appears to vary across brain regions [49] and depends on gephyrin and ankyrin/ spectrin supracomplexes [50], major genetic risk factors in a number of neuronal disorders including epilepsy [51]. Notably, the enrichment of  $\alpha 1$ -,  $\alpha 2$ -, and  $\alpha 3$ -GABAARs and their co-localization with gephyrin and/ or ankyrin shows strong regional differences [49] and appears to be mediated by independent and distinct receptor protein complexes, as the enrichment of one subunit was unaffected by the genomic deletion of the other [49]. Thus, indicating the presence of unique forms of GABAergic inhibitory transmission and hence addressable pharmacological profiles at the AIS. These could be exploited to guide existing treatments to the relevant patient cohorts and eventually result in the development of new personalized treatments [27,51].

### Targeting extrasynaptic GABA<sub>A</sub>R reserve pools

Extrasynaptic reserve pools represent an exciting key structure to expand GABAAR pharmacology beyond the postsynapse and the AIS. Postsynaptic localization of  $\alpha$ 5-GABA<sub>A</sub>Rs through gephyrin [52] is countered by extrasynaptic targeting through radixin [53]. Blockade of  $\alpha$ 5-GABARs accelerates reversal learning [54] and mice with a selective knockdown of  $\alpha$ 5-GABA<sub>A</sub>Rs in pyramidal neurons show improved spatial and trace fearconditioning memory [55]. Allosteric modulators with improved a5-GABAAR selectivity are being tested in neurodevelopmental disorders, depression, schizophrenia, and to improve cognitive function in neurodegenerative disorders and facilitate poststroke recovery [27] and are already being developed as photosensitive epilepsy medication [56]. Despite these impressive advances in targeting  $\alpha$ 5-GABA<sub>A</sub>Rs [57], the involved intracellular motifs and binding sites remain largely untapped. Insights into  $\alpha$ 5-GABA<sub>A</sub>R endogenous signalling will not only advance our understanding of memory and learning but could also contribute to the development of cognitive enhancers.

### GABA<sub>A</sub>R-associated proteins: challenges and opportunities

### Challenge: functional and physical mapping of intracellular GABA<sub>A</sub>R interactions

Targeting GABA<sub>A</sub>R-associated proteins requires precise knowledge about their distribution in specific brain areas, neuronal types and subregions, which, however, is still only rudimentary. The necessary functional and physical mapping of the involved intracellular GABA<sub>A</sub>R interaction surfaces demands the development and rigorous application of technologies with enhanced throughput and level of detail.

Mass spectrometric (MS) approaches reported numerous putative GABAAR-associated proteins [58]. As intracellular receptor interactions can be resolved and validated using short linear peptide fragments of the intracellular receptor regions [59], high-throughput MS approaches such as Protein Interaction Screen on Peptide Matrices [60] can be applied to resolve and map receptor subunit interactomes and their posttranslational regulation. The application of advanced methods such as protein cross-linking coupled with MS (XL-MS) [61], multiplexed proximity biotinylation [62], and their bioinformatic integration has been outlined recently [63]. These developments as well as long read sequencing will aid in grasping the complexity of gene regulation, specifically alternative splicing [64], and disease-relevant post-translational modifications at GABAergic synapses. Integration of experimental results into computational platforms, as recently provided by Lupascu et al., could allow to decipher how subcellular and intracellular pathways precisely affect individual GABAergic signalling events in physiology and pathology [65].

### Opportunity: short linear receptor motifs provide specificity

Characterization of a short list of GABAAR-associated proteins enabled the design of modulators of GABAergic signalling and postsynaptic labels [12] and recently studied GABAAR-associated proteins may be exploited in the same way (Table 1). Structural characterization, including X-ray crystallographic studies and recent cryoelectron tomograms [66], defined the intracellular GABAAR regions as intrinsically disordered, devoid of fixed secondary structural elements. By only adopting a fold on binding, short linear motifs within the intracellular receptor regions allow for specific interactions with more than one target. Such domain-motif interactions are observed across the proteome and are characterized by structural flexibility and tight post-translational control. This enables immediate adaption in response to changing stimuli, which may be a requirement for their role in structural postsynaptic plasticity.

### Opportunity: resolving the nanoscale arrangement of the inhibitory postsynapse

The inherently high binding energy densities of domain-motif interactions [12] suggest they could be harnessed not only as highly specific and potent inhibitors but also as compact fluorescent labels. GABA<sub>A</sub>R-associated protein targeting molecules may be turned into sophisticated tools for simultaneous visualization and manipulation of the neuronal system with molecular precision. When combined with fluorescent dyes and cell penetrating tags, they can be expected to visualize the nanometer-scaled machinery that underlies higher brain functions such as learning and memory, possibly even in living cells and tissue slices. As the fluorophore stays close to the target surface, they will provide the necessary nanometer resolution and localization precision. Moreover, highly affine and selective peptides could achieve stoichiometric labeling, enabling quantification of the target protein, and resolve the specific distribution of GABA<sub>A</sub>R subtypes and intracellular interactors [21,75]. When combined with UV-inducible photoinhibitors, they could provide photocontrol of GABA<sub>A</sub>R function *in situ* [67].

## A toolbox for exploring intracellular $GABA_AR$ interactions

A range of  $GABA_AR$  binding motifs was successfully transformed into bioactive actuators [12].  $GABA_AR$  modulation and visualization in brain slices has been

#### Figure 3



Toolbox for studying and targeting GABA<sub>A</sub>R-associated proteins. Newly available methods to empower molecular studies in high throughput and the development of pharmaceutical effectors acting on GABAAR-associated proteins. Binding sites and hierarchies that define GABAAR intracellular signalling are largely unknown. The depicted funnel symbolizes the striking discrepancy between numbers of candidate proteins and validated or characterized GABAAR-associated proteins. A major bottleneck is the elucidation of identified protein-protein interactions at molecular detail and affinity quantification. A conceptual workflow to achieve this task may include the screening of binding interfaces using synthetic biology approaches such as phage display, and microarrays obtained from parallel peptide synthesis. Newly reported methodological approaches could be applied to iteratively maturate peptide binders into bioactive GABAAR effectors or facilitate structural investigations to resolve the involved binding pocket. ECD: extracellular domain, TM: transmembrane domain.

achieved with peptides as well as the natural small molecule drug artemisinin [21-23]. Currently, the major bottleneck (Figure 3) in the development of such peptide-based effectors lies in the mapping and characterization of the involved interfaces that, until recently, necessitated laborious biophysical studies as outlined above [12].

Screening followed by iterative affinity maturation by sequence optimization and chemical modification in microarray format recently empowered the development of intracellularly active peptide-based modulators of GABAergic signalling [21]. Likewise, approaches combining large-scale phage display libraries with in silico modeling provided attractive starting points for the detailed characterization of GABAAR-associated protein interactions as recently exemplified by a study on 4.1 protein, erzin, moesin and radixin (FERM) domains [68]. The toolbox (Figure 3) for development of highaffinity peptide binders was furthermore expanded by nano- to picomolar scaled affinity determination of peptide-protein interactions in-solution [69] and size exclusion-based enrichment from pools of synthetic peptides combined with liquid chromatographytandem MS-based peptide sequencing [70]. Setups that combine miniaturized and automated peptide synthesis with high-throughput binding evaluation could be ideal to map and characterize intracellular GABAAR interactions and convert identified linear motifs into bioactive competitive binders (Figure 3).

### Conclusions

The pharmacological targeting of neurotransmittergated ion channel and G protein-coupled receptor associated proteins and protein complexes in the brain represents an emerging field with several compounds currently in clinical trials and first approved for clinical application [71]. In stark contrast, and despite the increasing number of newly identified transmembrane [73,74] and intracellularly GABAAR-associated proteins, the brain's natural inhibitory signalling pathways remain largely unexplored. Yet, in contrast to compounds that allosterically affect GABAARs across the brain, targeting the associated proteins has potential to adjust GABAergic activity with circuit precision, thereby potentially overcoming the severe side-effects of conventional GABAAR modulators and addressing unmet clinical need. To this end, the systematic application of novel approaches for the large-scale identification of GABAAR-associated proteins will not only determine their unique modes of action but also contribute to expanding GABAAR pharmacology.

### Author contributions

C.S. and H.M.M. wrote the manuscript and prepared the figures.

### Conflict of interest statement

C.S. and H.M.M. have no conflict of interest on this topic.

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#### References

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- \*\* of outstanding interest
- Oh WC, Smith KR: Activity-dependent development of 1. GABAergic synapses. Brain Res 2019, 1707:18-26.
- Barker JS, Hines RM: Regulation of GABAA receptor subunit 2. expression in substance use disorders. Int J Mol Sci 2020, 21: 4445
- З. Kim J, Son Y, Kim J, Lee S, Kang S, Park K, Kim SH, Kim JC, Kim J, Takayama C, et al.: Developmental and degenerative modulation of GABAergic transmission in the mouse hippocampus. Int J Dev Neurosci 2015, 47:320-332.
- Rajgor D, Purkey AM, Sanderson JL, Welle TM, Garcia JD, 4. Dell'Acqua ML, Smith KR: Local miRNA-dependent translational control of GABAAR synthesis during inhibitory longterm potentiation. Cell Rep 2020, 31:107785.
- Niwa F, Patrizio A, Triller A, Specht CG: cAMP-EPAC-Depen-dent regulation of gephyrin phosphorylation and GABAAR 5. trapping at inhibitory synapses. iScience 2019, 22:453-465.
- Maynard SA, Triller A: Inhibitory receptor diffusion dynamics. 6. Front Mol Neurosci 2019, 12:313.
- Pizzarelli R, Griguoli M, Zacchi P, Petrini EM, Barberis A, Cattaneo A, Cherubini E: Tuning GABAergic inhibition: 7. gephyrin molecular organization and functions. *Neuroscience* 2020, **439**:125–136.
- Humeau Y, Choquet D: The next generation of approaches to 8 investigate the link between synaptic plasticity and learning. Nat Neurosci 2019, 22:1536-1543.
- Yang X, Le Corronc H, Legendre P, Triller A, Specht CG: Dif-9. ferential homeostatic regulation of glycinergic and GABAergic nanocolumns at mixed inhibitory synapses. bioRxiv 2020. 2020.2011.2023.372383.
- 10. Crosby KC, Gookin SE, Garcia JD, Hahm KM, Dell'Acqua ML Smith KR: Nanoscale subsynaptic domains underlie the organization of the inhibitory synapse. Cell Rep 2019, 26. 3284-3297 e3283
- 11. Campbell BFN, Tyagarajan SK: Cellular mechanisms contrib-uting to the functional heterogeneity of GABAergic synapses. Front Mol Neurosci 2019. 12:187.
- 12. Khayenko V, Maric HM: Targeting GABAAR-associated proteins: new modulators, labels and concepts. Front Mol Neurosci 2019. 12.
- 13. Mele M, Costa RO, Duarte CB: Alterations in GABAA-receptor trafficking and synaptic dysfunction in brain disorders. Front Cell Neurosci 2019, 13:77.
- 14. Ali Rodriguez R, Joya C, Hines RM: Common ribs of inhibitory synaptic dysfunction in the umbrella of neurodevelopmental disorders. Front Mol Neurosci 2018, 11:132.
- 15. Gonzalez MI: Calpain-dependent cleavage of GABAergic proteins during epileptogenesis. Epilepsy Res 2019, 157: 106206.
- 16. Chiu CQ, Barberis A, Higley MJ: Preserving the balance: diverse forms of long-term GABAergic synaptic plasticity. Nat Rev Neurosci 2019, 20:272-281.

- 17. Matt L, Kim K, Chowdhury D, Hell JW: Role of palmitoylation of postsynaptic proteins in promoting synaptic plasticity. Front Mol Neurosci 2019, 12:8.
- 18. Shen ZC, Wu PF, Wang F, Xia ZX, Deng Q, Nie TL, Zhang SQ, Zheng HL, Liu WH, Lu JJ, *et al.*: Gephyrin palmitoylation in basolateral amygdala mediates the anxiolytic action of benzodiazepine. Biol Psychiatr 2019, 85:202-213.
- 19. Yang X, Specht CG: Subsynaptic domains in super-resolution microscopy: the treachery of images. Front Mol Neurosci 2019, 12:161.
- 20. Specht CG: Fractional occupancy of synaptic binding sites and the molecular plasticity of inhibitory synapses. Neuropharmacology 2020, 169, 107493.
- 21. Maric HM, Hausrat TJ, Neubert F, Dalby NO, Doose S, Sauer M, \* Kneussel M, Stromgaard K: Gephyrin-binding peptides visualize postsynaptic sites and modulate neurotransmission. Nat Chem Biol 2017, 13:153-160.

First example of a fundamentally novel approach for targeting GABAAR synaptic anchoring via gephyrin. First illustration of modulation of inhibitory receptors by interfering with the receptor/gephyrin interaction. Proof-of-concept demonstration that intracellular peptide labels provide superior tools to visualize inhibitory synapses in neurons, even at super-resolution

- 22. Kasaragod VB, Hausrat TJ, Schaefer N, Kuhn M, \*\* Christensen NR, Tessmer I, Maric HM, Madsen KL, Sotriffer C, Villmann C, et al.: Elucidating the molecular Basis for inhibitory neurotransmission regulation by artemisinins. Neuron 2019. 101. 673-689 e611.

First demonstration of modulation of GABAergic transmission via small drug-like compounds targeting the universal receptor binding pocket of the intracellular inhibitory scaffold protein gephyrin.

- Kittler JT, Chen G, Kukhtina V, Vahedi-Faridi A, Gu Z, Tretter V, Smith KR, McAinsh K, Arancibia-Carcamo IL, Saenger W, et al.: Regulation of synaptic inhibition by phospho-dependent binding of the AP2 complex to a YECL motif in the GABAA receptor gamma2 subunit. Proc Natl Acad Sci U S A 2008, 105: 3616-3621.
- Ge Y, Kang Y, Cassidy RM, Moon KM, Lewis R, Wong ROL, 24. Foster LJ, Craig AM: Clptm1 limits forward trafficking of GABAA receptors to scale inhibitory synaptic strength. Neuron 2018. 97:596-610 e598.
- 25. Halff EF, Szulc BR, Lesept F, Kittler JT: SNX27-Mediated recycling of neuroligin-2 regulates inhibitory signaling. Cell Rep 2019. 29. 2599-2607.e2596.

Halff et al. identify SNX27 as a novel mediator of neuroligin 2 surface directly influencing regulation, thereby number GABAergic transmission.

Davenport EC, Szulc BR, Drew J, Taylor J, Morgan T, Higgs NF, López-Doménech G, Kittler JT: Autism and schizophrenia-associated CYFIP1 regulates the balance of synaptic excita-26. tion and inhibition. Cell Rep 2019, 26. 2037-2051.e2036.

One representative example for several recent works on the complex role of CYFIP1/2 at the inhibitory synapse. The work of Davenport et al. highlights molecular details that underly its postsynaptic function.

- 27. Maramai S, Benchekroun M, Ward SE, Atack JR: Subtype selective gamma-aminobutyric acid type A receptor (GABAAR) modulators acting at the benzodiazepine binding site: an update. J Med Chem 2020, 63:3425-3446.
- 28. Olsen RW: GABAA receptor: positive and negative allosteric modulators. Neuropharmacology 2018, 136:10-22.
- Sieghart W, Savic MM: International union of basic and clinical pharmacology. CVI: GABAA receptor subtype- and function-selective ligands: key issues in translation to humans. 29. Pharmacol Rev 2018, 70:836-878.
- 30. Belelli D, Hogenkamp D, Gee KW, Lambert JJ: Realising the therapeutic potential of neuroactive steroid modulators of the GABAA receptor. Neurobiol Stress 2020, 12:100207.
- 31. Richter G, Liao VWY, Ahring PK, Chebib M: The Z-drugs zolpi-dem, zaleplon, and eszopiclone have varying actions on human GABA A receptors containing gamma1, gamma2, and gamma3 subunits. Front Neurosci 2020, 14:599812.

- 32. Silva J, Shao AS, Shen Y, Davies DL, Olsen RW, Holschneider DP, Shao XM, Liang J: Modulation of hippocampal GABAergic neurotransmission and gephyrin levels by dihydromyricetin improves anxiety. *Front Pharmacol* 2020, 11: 1008.
- Parakala ML, Zhang Y, Modgil A, Chadchankar J, Vien TN,
   Ackley MA, Doherty JJ, Davies PA, Moss SJ: Metabotropic, but not allosteric, effects of neurosteroids on GABAergic inhibition depend on the phosphorylation of GABAA receptors. *J Biol Chem* 2019, 294:12220–12230.

This study demonstrates the critical contribution of intracellular  $GABA_AR$  signalling to the long lasting beneficial therapeuthic effects of neurosteroids.

- Althaus AL, Ackley MA, Belfort GM, Gee SM, Dai J, Nguyen DP, Kazdoba TM, Modgil A, Davies PA, Moss SJ, et al.: Preclinical characterization of zuranolone (SAGE-217), a selective neuroactive steroid GABAA receptor positive allosteric modulator. Neuropharmacology 2020, 181:108333.
- Kralic JE, Sidler C, Parpan F, Homanics GE, Morrow AL, Fritschy JM: Compensatory alteration of inhibitory synaptic circuits in cerebellum and thalamus of gamma-aminobutyric acid type A receptor alpha1 subunit knockout mice. J Comp Neurol 2006, 495:408–421.
- Mondoloni S, Durand-de Cuttoli R, Mourot A: Cell-specific neuropharmacology. Trends Pharmacol Sci 2019, 40:696–710.
- Lorenz-Guertin JM, Bambino MJ, Das S, Weintraub ST, Jacob TC: Diazepam accelerates GABAAR synaptic exchange and alters intracellular trafficking. Front Cell Neurosci 2019, 13:163.
- Schmitz A: Benzodiazepines: the time for systematic change is now. Addiction 2020, 116:219–221.
- Shi YW, Zhang Q, Cai K, Poliquin S, Shen W, Winters N, Yi YH, Wang J, Hu N, Macdonald RL, et al.: Synaptic clustering differences due to different GABRB3 mutations cause variable epilepsy syndromes. Brain 2019, 142:3028–3044.
- Contreras A, Hines DJ, Hines RM: Molecular specialization of GABAergic synapses on the soma and axon in cortical and hippocampal circuit function and dysfunction. Front Mol Neurosci 2019, 12:154.
- 41. Maric HM, Mukherjee J, Tretter V, Moss SJ, Schindelin H: Gephyrin-mediated gamma-aminobutyric acid type A and glycine receptor clustering relies on a common binding site. *J Biol Chem* 2011, 286:42105–42114.
- Bai G, Wang Y, Zhang M: Gephyrin-mediated formation of inhibitory postsynaptic density sheet via phase separation. *Cell Res* 2020, 0:1–14.

This article provides first evidence that the formation of the inhibitory postsynaptic density (PSD) is, similar to the excitatory PSD, governed by phase separation-mediated condensation of scaffold protein/ neurotransmitter receptor complexes and further reveals post-translational regulation pathways.

- 43. Maric HM, Kasaragod VB, Haugaard-Kedstrom L, Hausrat TJ, Kneussel M, Schindelin H, Stromgaard K: Design and synthesis of high-affinity dimeric inhibitors targeting the interactions between gephyrin and inhibitory neurotransmitter receptors. Angew Chem Int Ed Engl 2015, 54:490–494.
- George S, Bear Jr J, Taylor MJ, Kanamalla K, Fekete CD, Chiou TT, Miralles CP, Papadopoulos T, De Blas AL: Collybistin SH3-protein isoforms are expressed in the rat brain promoting gephyrin and GABA-A receptor clustering at GABAergic synapses. J Neurochemistry 2020, 00:1–20.
- Hines RM, Maric HM, Hines DJ, Modgil A, Panzanelli P,
   Nakamura Y, Nathanson AJ, Cross A, Deeb T, Brandon NJ, et al.: Developmental seizures and mortality result from reducing GABAA receptor alpha2-subunit interaction with collybistin. Nat Commun 2018, 9:3130.

This work identified and quantified the direct interaction between collybistin and  $\alpha$ 2-GABA<sub>A</sub>Rs and demonstrates the relevance of the identified motif for maintaining  $\alpha$ 2-GABA<sub>A</sub>Rs at the AIS and provides a mouse model for eppilepsy disorder.

**46.** Nathanson AJ, Zhang Y, Smalley JL, Ollerhead TA, Rodriguez \*\* Santos MA, Andrews PM, Wobst HJ, Moore YE, Brandon NJ, Hines RM, *et al.*: Identification of a core Amino acid motif within the alpha subunit of GABAARs that Promotes inhibitory Synaptogenesis and resilience to seizures. *Cell Rep* 2019, 28. 670-681 e678.

This article highlights the unique role of  $\alpha$ 2-GABA<sub>A</sub>Rs at the AIS for seizure resillience and indicates utility of  $\alpha$ 2/ $\alpha$ 3 specific benzodiazepines in a subgroup of eppilepsy patients. First proof of concept that neuronal dysfunctions resembling the pathology of human eppilepsy can be reconsituted by manipulation of the intracellular receptor complexes.

- Zou G, Chen Q, Chen K, Zuo X, Ge Y, Hou Y, Pan T, Pan H, Liu D, Zhang L, *et al.*: Human hyperekplexic mutations in Glycine receptors disinhibit the brainstem by Hijacking GABAA receptors. *iScience* 2019, 19:634–646.
- 48. Takano T, Wallace JT, Baldwin KT, Purkey AM, Uezu A,
  Courtland JL, Soderblom EJ, Shimogori T, Maness PF, Eroglu C, et al.: Chemico-genetic discovery of astrocytic control of inhibition in vivo. Nature 2020, 588:296–302.

Takano et al. present a working model for the interaction of astrocytes with neurons on proteomic level and their influence on GABAergic synapse formation and function.

- Gao Y, Heldt SA: Enrichment of GABAA receptor alphasubunits on the axonal initial segment shows regional differences. Front Cell Neurosci 2016, 10:39.
- Nathanson AJ, Davies PA, Moss SJ: Inhibitory synapse formation at the axon initial segment. Front Mol Neurosci 2019, 12:266.
- Maljevic S, Moller RS, Reid CA, Perez-Palma E, Lal D, May P, Lerche H: Spectrum of GABAA receptor variants in epilepsy. *Curr Opin Neurol* 2019, 32:183–190.
- Brady ML, Jacob TC: Synaptic localization of alpha5 GABA (A) receptors via gephyrin interaction regulates dendritic outgrowth and spine maturation. *Dev Neurobiol* 2015, 75: 1241–1251.
- Hausrat TJ, Muhia M, Gerrow K, Thomas P, Hirdes W, Tsukita S, Heisler FF, Herich L, Dubroqua S, Breiden P, et al.: Radixin regulates synaptic GABAA receptor density and is essential for reversal learning and short-term memory. Nat Commun 2015, 6:6872.
- 54. Davenport CM, Rajappa R, Katchan L, Taylor CR, Tsai MC,
   \*\* Smith CM, de Jong JW, Arnold DB, Lammel S, Kramer RH: Relocation of an extrasynaptic GABAA receptor to inhibitory synapses Freezes excitatory synaptic strength and Preserves memory. Neuron 2021, 109. 123-134 e124.
   This work presents a model of α5-GABA<sub>A</sub>R-mediated synaptic plas-

This work presents a model of  $\alpha$ 5-GABA<sub>A</sub>R-mediated synaptic plasticity that controls the preservation of learned associations in mice and has strong implications for  $\alpha$ 5-GABA<sub>A</sub>R-targeted therapy.

- Engin E, Sigal M, Benke D, Zeller A, Rudolph U: Bidirectional regulation of distinct memory domains by alpha5-subunitcontaining GABAA receptors in CA1 pyramidal neurons. *Learn Mem* 2020, 27:423–428.
- Gurrell R, Gorman D, Whitlock M, Ogden A, Reynolds DS, DiVentura B, Abou-Khalil B, Gelfand M, Pollard J, Hogan RE, et al.: Photosensitive epilepsy: robust clinical efficacy of a selective GABA potentiator. Neurology 2019, 92:e1786–e1795.
- Jacob TC: Neurobiology and therapeutic potential of alpha5-GABA type A receptors. Front Mol Neurosci 2019, 12:179.
- Murtaza N, Uy J, Singh KK: Emerging proteomic approaches to identify the underlying pathophysiology of neurodevelopmental and neurodegenerative disorders. *Mol Autism* 2020, 11:27.
- Langlhofer G, Schaefer N, Maric HM, Keramidas A, Zhang Y, Baumann P, Blum R, Breitinger U, Stromgaard K, Schlosser A, et al.: A novel glycine receptor variant with startle disease affects syndapin I and glycinergic inhibition. J Neurosci 2020, 40:4954–4969.
- Dittmar G, Hernandez DP, Kowenz-Leutz E, Kirchner M, Kahlert G, Wesolowski R, Baum K, Knoblich M, Hofstatter M, Muller A, et al.: PRISMA: protein interaction screen on peptide matrix reveals interaction footprints and modificationsdependent interactome of intrinsically disordered C/EBPbeta. *iScience* 2019, 13:351–370.

 Gonzalez-Lozano MA, Koopmans F, Sullivan PF, Protze J,
 Krause G, Verhage M, Li KW, Liu F, Smit AB: Stitching the synapse: cross-linking mass spectrometry into resolving synaptic protein interactions. Sci Adv 2020, 6, eaax5783.

Employing XL-MS, an extensive dataset of synaptic protein-protein interactions, including specific interaction sites, was generated. This or similar approaches are yet to be applied on  $GABA_AR$ -associated proteins.

- Hamdan H, Lim BC, Torii T, Joshi A, Konning M, Smith C, Palmer DJ, Ng P, Leterrier C, Oses-Prieto JA, *et al.*: Mapping axon initial segment structure and function by multiplexed proximity biotinylation. *Nat Commun* 2020, 11:100.
- 63. Koopmans F, van Nierop P, Andres-Alonso M, Byrnes A, Cijsouw T, Coba MP, Cornelisse LN, Farrell RJ, Goldschmidt HL, Howrigan DP, *et al.*: SynGO: an evidence-based, expertcurated knowledge base for the synapse. *Neuron* 2019, 103. 217-234 e214.
- Furlanis E, Scheiffele P: Regulation of neuronal differentiation, function, and plasticity by alternative splicing. Annu Rev Cell Dev Biol 2018, 34:451–469.
- Lupascu CA, Morabito A, Ruggeri F, Parisi C, Pimpinella D,
   Pizzarelli R, Meli G, Marinelli S, Cherubini E, Cattaneo A, et al.: Computational modeling of inhibitory transsynaptic signaling in hippocampal and cortical neurons expressing Intrabodies against gephyrin. Front Cell Neurosci 2020, 14:173.

Lupascu at al. provide a computational plattform to decipher inhibitory synaptic transmission upon manipulation of the inhibitory scaffold protein gephyrin, by integrating complementary approaches.

- Liu YT, Tao CL, Zhang X, Xia W, Shi DQ, Qi L, Xu C, Sun R, Li XW, Lau PM, et al.: Mesophasic organization of GABAA receptors in hippocampal inhibitory synapses. Nat Neurosci 2020, 23:1589–1596.
- Mortensen M, Krall J, Kongstad KT, Brygger BM, Lenzi O, Francotte P, Sorensen TE, Nielsen B, Jensen AA, Smart TG, et al.: Developing new 4-PIOL and 4-PHP analogues for photoinactivation of gamma-aminobutyric acid type A receptors. ACS Chem Neurosci 2019, 10:4669–4684.

- Ali M, Khramushin A, Yadav VK, Schueler-Furman O, Ivarsson Y: Defining binding motifs and dynamics of the multi-pocket FERM domain from ezrin, radixin, moesin and merlin. *bioRxiv* 2020. 2020.2011.2023.394106.
- Schulte C, Khayenko V, Nordblom NF, Tippel F, Peck V,
   Gupta AJ, Maric HM: High-throughput determination of protein affinities using unmodified peptide libraries in nanomolar scale. *iScience* 2021, 24:101898.

This work demonstrates for the first time the application of temperature related intensity change (TRIC)-based measurements for the affinity determination of protein-peptide interactions. As a proof-of concept, gephyrin/GABA<sub>A</sub>R subunit interactions are quantified in solution. In addition, receptor binding sites of native GABA<sub>A</sub>R-associated proteins within whole brain homegenates are precisely mapped and quantified.

- Touti F, Gates ZP, Bandyopadhyay A, Lautrette G, Pentelute BL: In-solution enrichment identifies peptide inhibitors of protein-protein interactions. Nat Chem Biol 2019, 15:410–418.
- Rosenbaum MI, Clemmensen LS, Bredt DS, Bettler B, Stromgaard K: Targeting receptor complexes: a new dimension in drug discovery. Nat Rev Drug Discov 2020, 19:884–901.
- 72. Smith KR, Muir J, Rao Y, Browarski M, Gruenig MC, Sheehan DF, Haucke V, Kittler JT: Stabilization of GABA(A) receptors at endocytic zones is mediated by an AP2 binding motif within the GABA(A) receptor beta3 subunit. J Neurosci 2012, 32: 2485–2498.
- Wenyan H, Shepard RD, Wei L: Regulation of GABAARs by transmembrane accessory proteins. *Trends Neurosci* 2021, 44:152–165.
- 74. Castellano D, Shepard RD, Wei L: Looking for novelty in an "old" receptor: recent advances toward our understanding of gabaars and their implications in receptor pharmacology. *Front Neurosci* 2021, 14:1420.
- Wang L, Tran M, D'Este E, Roberti J, Koch B, Xue L, Johnsson K: A general strategy to develop cell permeable and fluorogenic probes for multicolour nanoscopy. Nat Chem 2020, 12: 165–172.